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Herbivory on freshwater and marine macrophytes: a review and perspective

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writing of the manuscript

Abstract

Until the 1990s, herbivory on aquatic vascular plants was considered to be of minor importance, and the predominant view was that freshwater and marine macrophytes did not take part in the food web: their primary fate was the detritivorous pathway. In the last 25 years, a substantial body of evidence has developed that shows that herbivory is an important factor in the ecology of vascular macrophytes across freshwater and marine habitats. Herbivores remove on average 40-48% of plant biomass in freshwater and marine ecosystems, which is typically 5-10 times greater than reported for terrestrial ecosystems. This may be explained by the lower C:N stoichiometry found in submerged plants. Herbivores affect plant abundance and species composition by grazing and bioturbation and therewith alter the functioning of aquatic ecosystems, including biogeochemical cycling, carbon stocks and primary production, transport of nutrients and propagules across ecosystem boundaries, habitat for other organisms and the level of shoreline protection by macrophyte beds. With ongoing global environmental change, herbivore impacts are predicted to increase. There are pressing needs to improve our management of undesirable herbivore impacts on macrophytes (e.g. leading to an ecosystem collapse), and the conflicts between people associated with the impacts of charismatic mega-herbivores. While simultaneously, the long-term future of maintaining both viable herbivore populations and plant beds should be addressed, as both belong in complete ecosystems and have co-evolved in these long before the increasing influence of man. Better integration of the freshwater, marine, and terrestrial herbivory literatures would greatly benefit future research efforts.

Keywords: climate change, conservation, ecosystem functions, grazing, seagrass, stoichiometry

1. Introduction: 25 years of research on herbivory on macrophytes

1.1 *Setting the scene*

In the 1990s two seminal papers appeared in *Aquatic Botany* that urged for a complete change in the paradigm that had been dominating macrophyte ecology. Despite some early work on the impact of waterbirds on freshwater and marine angiosperms (Jupp and Spence, 1977; Jacobs et al., 1981), until then, herbivory on aquatic vascular plants was considered to be of minor importance, and the predominant view was that freshwater and marine macrophytes did not take part in the food web (e.g. Shelford (1918) and their primary fate was the detritivorous pathway (Polunin, 1984; Duarte and Cebrian, 1996). But in 1991, Lodge argued that, contrary to conventional wisdom, live freshwater macrophytes are engaged in aquatic food webs. In 1998, Cebrian and Duarte highlighted that, while seagrasses suffered modest herbivory rates on average, such rates were highly variable, and the importance of seagrass-herbivore interactions should not be discounted. Following on from these two papers, Lodge (1998) provided further evidence for the important role of herbivores in freshwater habitats, as compared to other biomes; and Valentine and Heck (1999) demonstrated that grazing on seagrasses is widespread in the world's oceans.

Together, these landmark papers put macrophyte herbivory on the map. Since then, there has been a strong increase in the amount of studies that investigated herbivory on freshwater macrophytes and seagrasses. In this study, we review what we have learned in the 25 years that followed the appearance of Lodge (1991). Furthermore, we identify new topics that have emerged over this time. These new topics include the fast changes that may occur in macrophyte-herbivore relationships with the ongoing global environmental change, as well as the potential conflicts between herbivore conservation and herbivore impacts on aquatic ecosystems. Finally, we discuss how we can improve our understanding of herbivore impacts and what tools may help us in

achieving this. Following the approach of the seminal papers listed above, we focus primarily on aquatic angiosperms (submerged, floating and emergent) and address both freshwater and marine ecosystems.

1.2 Why thinking about herbivory on macrophytes has changed over the last 25 years

The paradigm shift in our perception of macrophyte herbivory, from being considered negligible to being acknowledged as a key factor shaping benthic ecosystems, is not only caused by an increase in scientific interest fostered by these landmark papers: the effect of herbivory became also more conspicuous over the last 25 years. The reasons for this are methodological, anthropogenic and ecological.

Methodological improvements for estimating herbivory included observation methods, such as bite mark counts (Cebrian and Duarte, 1998), experimental approaches, such as herbivore exclusions (see Poore et al. (2012) and Wood et al. (2016) for syntheses of marine and freshwater habitats, including macro-algae) and direct methods, including video bite counts or isotopic signatures (see Table 4 for details).

Anthropogenic effects included increases in the densities of aquatic and marine herbivores as a result of increased protection, predator removal, food subsidies from agriculture, and the introduction of exotic herbivores (Estes et al., 2011). For example, steep increases in herbivory rates have been reported for sea turtles in the Arabian Sea and Indonesia (Kelkar et al., 2013b, a), (Christianen et al., 2014), for herbivorous fish in the Mediterranean (Pages et al., 2012) and for geese in Northwestern Europe and North America (Jefferies et al., 2003; Van Eerden et al., 2005). However, it should be noted that despite recent local increases in herbivory, which have attracted attention to the role of herbivores in benthic ecosystems, over longer time frames in particularly species of large herbivores have experienced strong global declines (Jackson, 1997; McCauley et al., 2015; Bakker et al. 2016b).

Furthermore, the recent spread of exotic herbivores had major consequences for macrophyte establishment and survival in many areas worldwide. For example, tropical lessepsian rabbitfishes (*Siganus spp.*) cause overgrazing of macroalgae and seagrasses at the Eastern Mediterranean (Verges et al., 2014b), chubs and rabbit fishes (*Kyphosus spp.* and *Siganus spp.*, Siganidae) overgraze Australian and Japanese kelp forests (Verges et al., 2014a), North-American red-swamp crayfish (*Procambarus clarkii*) have depleted submerged plant meadows in shallow lakes across Europe (Rodriguez et al., 2003; Gherardi and Acquistapace, 2007; van der Wal et al., 2013), and intentional introductions of grass carp (*Ctenopharyngodon idella*) have been considered a threat to native macrophytes (Wittmann et al., 2014).

Ecological effects are related to the oligotrophication of many European freshwater systems, which resulted in their re-colonization with submerged macrophytes (Jeppesen et al., 2005). In many systems, however, the impact of aquatic herbivores sufficed to halt or reverse such recolonization (Körner and Dugdale, 2003; Hilt, 2006; Bakker et al., 2013; Hilt et al., 2013; Eigemann et al., 2016).

2. Quantitative impacts of herbivores in aquatic systems

2.1. Quantitative impact of herbivores on plant biomass across ecosystems

A growing body of primary research has demonstrated herbivore-induced changes in one or more measures of macrophyte abundance, including biomass, two-dimensional cover, volume, and individual density (Kirsch et al., 2002; Marklund et al., 2002; Tomas et al., 2005; Prado et al., 2007; Christianen et al., 2012; Pages et al., 2012; Wood et al., 2012a; Kelkar et al., 2013b, a; Christianen et al., 2014; Bakker et al., 2016b). These studies, synthesized in several reviews (Cyr and Pace, 1993; Valentine and Duffy, 2006; Gruner et al., 2008; Poore et al., 2012), confirmed herbivores as key drivers of benthic ecosystems around the world. The overwhelming majority of studies reported a reduction in macrophyte abundance as a result of herbivory. Indeed, a recent meta-analysis of 326

experiments in which freshwater herbivores were excluded found that herbivory reduced macrophyte biomass by 47.2 ± 3.4 % (average \pm CI) (Wood et al., 2016). Of these, 300 experiments reported a reduction in macrophyte biomass, while 26 experiments reported positive effects or no changes. Similarly, a meta-analysis on grazing impacts on marine macrophytes found that herbivores reduce macrophyte abundance (both submerged angiosperms and macro-algae) by 68% on average (Poore et al., 2012).

Despite their historical disregard, the removal of vascular plant biomass by herbivores is, on average, much larger in aquatic than in terrestrial ecosystems. The most recent meta-analyses available for terrestrial, freshwater and marine habitats (Turcotte et al., 2014; Wood et al., 2016) show that median biomass removal by herbivores is 4-8 % in terrestrial ecosystems, while it is 44-48% in freshwater and 40-44% in marine ecosystems (Fig 1a-c). Thus, herbivores remove on average 5-10 times more vascular plant biomass in aquatic ecosystems than in terrestrial ones.

Yet, the impact of herbivores on vascular plant biomass removal is much more variable in aquatic than in terrestrial ecosystems, and it ranges as broadly as between 0 and 100% of biomass removal (Fig 1a-c). Underlying explanations for the large range of herbivore effects in aquatic ecosystems are still unknown. Potential mechanisms involve bottom-up effects, such as variation in plant productivity, nutritional quality, stoichiometry, resistance and tolerance to grazing (Cebrian et al., 2009); and top-down effects, such as variation in herbivore abundance, feeding efficiency, size, taxonomy, mobility, metabolism and predator effects (Borer et al., 2005).

2.2. Bottom-up effects: the plant's perspective

2.2.1. Primary productivity and herbivory rates

Studies in aquatic systems traditionally focussed on primary production of phytoplankton in pelagic habitats, and only recently littoral areas received more attention (Vadeboncoeur et al., 2002;

Brothers et al., 2013). Theoretical predictions based on Lotka-Volterra models suggest that grazing should increase with primary productivity (Gruner et al., 2008), because as plants produce more tissues or do so at faster rates, herbivores can increase their rates of consumption. This prediction relies in the assumption that herbivory rates are bottom-up regulated by the availability of plant tissues. Empirical evidence is however conflicting. A meta-analysis by Cyr and Pace (1993) concluded that herbivory increases with primary production in both terrestrial and aquatic systems. However, a more recent meta-analysis found no significant effect of productivity on herbivory (or interaction strength) in aquatic systems (e.g. (Borer et al., 2005)). In fact, it is widely recognized that food quality and defences also have major effects on herbivore performance that might mask any herbivory-productivity relationship. Using a plant growth model, Hidding et al. (2016) suggested that herbivore grazing effects on macrophytes become important above certain thresholds in periphyton shading and thus reduced production of plants.

2.2.2. *Plant stoichiometry*

Differences in herbivory rates across plant taxa have also been attributed to differences in plant quality, as perceived by herbivores (Lodge, 1991; Cronin et al., 2002). Generally, there is a positive relationship between the nitrogen content in the plant's tissue and its consumption by herbivores (Cebrian and Lartigue, 2004). This pattern holds both within and across ecosystems (Elser et al., 2000; Cebrian and Lartigue, 2004; Cebrian et al., 2009). Hence, it has been suggested that higher herbivore consumption rates in aquatic plants might be explained by their higher quality as food – arising from the lack of carbon-rich structural compounds that strengthen cell walls, increase resistance to herbivores, and reduce digestibility in terrestrial plants (Gruner et al., 2008). Unfortunately, most comparisons between aquatic and terrestrial systems undertaken to date restricted their estimates of aquatic herbivory to phytoplankton consumption, thus giving limited or no information on the consumption of aquatic macrophytes (Elser et al., 2000). Comparing herbivory

rates on, and food quality (e.g., nitrogen content) of, aquatic and terrestrial vascular plants would be more informative, because they have a close phylogenetic affinity and most differences could be attributed to the ecosystems they inhabit (Hay, 1991; Gross and Bakker, 2012; Burkepile, 2013).

A compilation of data on C:N ratios of vascular plants in terrestrial, freshwater and marine ecosystems reveals that they differ strongly between these systems (Fig 1d-f). Median C:N ratio decreased strongly from terrestrial vascular plants (25-30) and marine macrophytes (24-28) to freshwater macrophytes (12-16) (Fig 1d-f). The high N content (thus low C:N ratio) of freshwater macrophytes may indicate a higher quality to herbivores, potentially explaining the high herbivory rates found in freshwater systems. Taking into account the variation in growth forms that occurs in aquatic ecosystems makes this pattern stronger (Fig 2a-d). Herbivory rates are lower on emergent macrophytes (median freshwater = 36-48%; median marine = 24-36%) than on submerged macrophytes (median freshwater and marine = 48-60%) in both freshwater and marine ecosystems (Fig 2a,b). These differences fit closely the C:N ratio of the different plant growth forms, as emergent macrophytes have higher C:N ratios (median freshwater = 28-32; median marine = 24-28) than submerged macrophytes (median freshwater = 8-12; median marine = 20-24), particularly in freshwater ecosystems (Fig 2c,d).

Our data compilation indicates that differences in plant quality between terrestrial plants and macrophytes show close links to variation in herbivory rates. Submerged plants need less structural components, resulting in higher N content and higher rates of herbivory. In contrast, herbivory rates and N content of emergent macrophytes are similar to those found in terrestrial plants (Fig 1a, 1d, 2a-d). Although N content is acknowledged as an important determinant of plant quality to herbivores, other nutrients and secondary compounds may also influence it. For example, freshwater macrophytes and seagrasses have anti-herbivore defenses (Verges et al., 2008; Gross and Bakker, 2012), which may cause low palatability - even in highly nutritious species. Phenolics, terpenoids and nitrogenated compounds can be efficient deterrents of aquatic herbivores, although

deterrence of generalist herbivores often comes at the cost of higher preference by specialised consumers (Verges et al., 2007). Interestingly, in freshwater systems, the concentrations of phenolic compounds decrease from emergent to floating to submerged plants (Smolders et al., 2000), suggesting not only higher nutritional quality, but also less defended tissues, resulting in generally higher palatability of submerged plants. However, this issue still deserves more research, since most work has focused on macro-algae (e.g. (Hay, 1996)). Besides the differences between ecosystems discussed above, spatial and temporal variation in herbivory at population and individual-plant level has also been linked to plant quality (C:N, % N) and to the presence of anti-herbivore defences (Hacker and Bertness, 1995; Preen, 1995; Verges et al., 2007; Prado et al., 2010).

2.3. Top-down effects: the herbivore's perspective

2.3.1. Herbivore density

Within individual herbivore species, herbivore impact on aquatic plant abundance is positively related to herbivore density (Stott and Robson, 1970; Valentine and Heck, 1991; Wood et al., 2012a; Kelkar et al., 2013b). However, analyses of herbivore impacts across multiple herbivore species have found no relationship between herbivore density and macrophyte abundance, probably due to the confounding effects of interspecific differences in herbivore ecology (Marklund et al., 2002; Wood et al., 2012a). For example, among plant-eating waterfowl, substantial differences exist in mean adult body mass (from the 24 g of ocellated crake, *Micropygia schomburgkii*, to the 12,000 g of trumpeter swan, *Cygnus buccinator*), which is known to influence the species' diet and the absolute quantity of vegetation consumed (Wood et al., 2012a). Consequently, when herbivore densities were estimated as biomass densities (thus accounting for interspecific differences in individual body mass), a significant negative relationship between herbivore biomass density and macrophyte abundance was detected (Wood et al., 2012a; Wood et al., 2016). Only at relatively low herbivore densities were

positive changes in plant abundance reported, suggesting that in aquatic systems greater herbivore densities overwhelm plant compensatory growth responses (Wood et al., 2016).

2.3.2. Size, mobility and taxonomy of herbivores

By virtue of their size, large terrestrial herbivores are critical agents of change and maintenance of the ecosystems they inhabit (Owen-Smith, 1988; Bakker et al., 2016a). In aquatic environments, large herbivores have also been identified as key species (McCauley et al., 2015), often considered ecosystem engineers (e.g. (Bakker et al., 2016b)). It is not surprising, thus, that a recent meta-analysis found significantly stronger impacts of macrograzers (fishes, urchins and large molluscs) than mesograzers (amphipods, isopods and small molluscs) on marine macrophytes (algae, seagrasses and salt marshes) (Poore et al., 2012).

Another recent meta-analysis by Wood et al (2016) found substantial between-taxa differences in effects of herbivores on the abundance of freshwater and marine macrophytes. Echinoderms, molluscs, and fish had relatively large impacts on plants, while insects and birds had relatively low impacts. The reason for these differences may be the mobility and habitat preferences of each of these groups. Fully aquatic species that live underwater permanently, have been shown to produce the greatest impacts on aquatic plants (Bakker et al., 2016b), while facultative aquatic grazers, such as insects and birds, spread their activity more between different ecosystems (thanks to their high mobility), thus spreading also their impact between such ecosystems. Wood et al. (2016) also pointed out the fact that some herbivores (such as echinoderms, crayfish and molluscs) have restricted mobility and produce intense impacts due to their bulk grazing strategies, since they consume multiple tissues types and species, thus affecting a greater proportion of a plant community (Lodge et al., 1998).

2.3.3. *Herbivory and omnivory*

Most animals that consume aquatic plants are omnivores, very few are strict herbivores. There is a positive relationship between body size and degree of herbivory in aquatic omnivores: the larger the consumer, the more important plant consumption (granivory and folivory, particularly the latter) is in their diet (Clements et al., 2009; Wood et al., 2012a). Hence, per capita, large herbivores have the largest impact on plant abundance through grazing. The high level of generalistic feeding and omnivory of aquatic plant consumers may be related to the higher impact of herbivory on aquatic plants, because it may relax the effect of direct density dependence on consumers: whenever consumers overgraze plants, they can switch to alternative food sources (algae, detritus or animal prey) within or nearby the water body (Grey and Jackson, 2012). Furthermore, in aquatic systems, large herbivores and omnivores often feed on both aboveground and belowground plant material, which may multiply their impact on plants, due to the depletion of underground plant resources for regrowth.

Herbivore impact may also be enhanced by non-consumptive effects, which are documented to be severe in freshwater and marine macrophytes, e.g. bioturbation (Lodge, 1991). By initiating bare patches, herbivores can create focal points for further erosion of macrophyte meadows by waves in shallow areas (Christianen et al., 2013). Trampling, fecal deposition and increasing nutrient concentrations may also play a role. Whilst previous authors have cautioned against the assumption that herbivore effects on plants represent exclusively grazing losses due to consumption (Mitchell and Wass, 1996b), many studies continue to ignore the roles of non-consumptive effects.

2.4. *Latitude*

Potential variance in herbivore impacts on plant abundance across different latitudes has been a topic of growing interest among ecologists. However, the evidence for a role of latitudinal effects is

limited. For example, in macrophyte feeding assays carried out by Morrison and Hay (2012), only one out of three crayfish species showed a preference associated with latitude (a preference for higher latitude plants). Schemske et al. (2009) argued that, across all ecosystems, herbivore impacts are greater at lower latitudes. In contrast, a meta-analysis by Gruner et al. (2008) reported an increase in herbivore effects at higher latitudes in freshwater ecosystems, but not in marine ecosystems. Three more recent meta-analyses with larger sample sizes found no evidence of latitudinal gradients in herbivore impacts on aquatic macrophyte abundances, some including both vascular plants and macro-algae (Moles et al., 2011; Poore et al., 2012; Wood et al., 2016).

3. Ecosystem consequences of herbivory

With the establishment of herbivory as an important factor regulating plant abundance, Lodge (1991) ended his review with the conclusion that “the functional importance of grazing remains largely untested”. Indeed, this has been an emerging field of research over the last 25 years, with particular progress over the last 5 years. By their presence, herbivores may induce direct changes in plant dynamics and indirect effects on ecosystem functioning (Fig. 3). The effects of marine herbivores may include stimulated production of seagrass (Valentine et al., 1997; Moran and Bjorndal, 2005; Vonk et al., 2008b; Christianen et al., 2012), changes in seagrass meadow structure (Lal et al., 2010), and the reduction of the flux of organic matter and nutrients to sediments and plants (by short circuiting the detrital cycle; (Thayer et al., 1982; Vonk et al., 2008a) or their export to nearby habitats (Christianen et al., 2012). In salt marshes and aquatic ecosystems, an additional effect of herbivores is the return of nutrients through faeces and urine (Bazely and Jefferies, 1985; Hik et al., 1991; Frank et al., 2000), though in seagrass grazed by turtles this effect is reduced by nutrient transport to turtle resting areas (Christianen et al., 2012).

3.1 *Plant abundance and species composition and diversity*

While the largely negative impacts of herbivores on macrophyte abundance have been well documented through short-term enclosure studies, the persistence of such effects it is often less clear. In particular, highly-mobile herbivores such as waterfowl can cause large reductions in plant abundance before switching to ungrazed sites (Wood et al., 2012b). Ecosystem responses to fluctuating grazing pressure have received little attention to date. However, there is some evidence that repeated episodes of grazing over time, such as by breeding colonies of snow geese (*Chen caerulescens*) in wetlands, can cause sustained long-term shifts in species composition and declines in plant abundance (Kerbes et al., 1990). Where non-selective, generalist herbivores feed on mixed assemblages of macrophyte, herbivores can increase species evenness by reducing the abundance of dominant competitors relative to sub-dominant macrophyte species (Hidding et al., 2010b; Wood et al., 2012b). However, when they favor certain subordinate plant species over the dominant, they can reduce evenness (Hidding et al. 2010a).

Herbivory in tropical seagrasses (e.g. by sea urchins and green turtles) can influence species composition (Vonk et al., 2008b; Kelkar et al., 2013b; Hernandez and van Tussenbroek, 2014), but often in contrasting ways. In tropical multispecies meadows, slow growing climax seagrass species (*Thalassia hemprichii*) were promoted by small herbivores (urchins), while in more intensively grazed meadows species difference in herbivore's grazing preferences resulted in the dominance of fast growing pioneer species (Kelkar et al., 2013b). Larger grazers that consume belowground plant parts and create bare sediment patches, such as green turtles and dugongs, also introduce species heterogeneity by setting back species succession in grazed plots (Aragones et al., 2006; Christianen et al., 2013).

3.2 *Ecosystem functions and services of aquatic herbivores*

3.2.1. *Seed and propagule dispersal*

Besides their direct effects on their food plants, aquatic herbivores provide a key service for aquatic ecosystems: the passive dispersal of a broad variety of aquatic organisms, including the aforementioned food plants, as well as many other taxa attached or associated to them (Figuerola and Green, 2002a; Brochet et al., 2010a; van Leeuwen et al., 2012). Dispersal by most inland-water herbivores contributes to the redistribution of individuals within single wetlands and among nearby ones; while waterbirds (most notably, migratory species) are the main vector of long-distance dispersal among wetlands situated at separated watersheds, from regional to continental scales (Viana et al., 2013b; Viana et al., in press). Among waterbirds, the frequency and scale of long-distance dispersal events is known to vary with the vector's morphology, to depend on the migratory strategy and to scale negative with body mass (Green and Figuerola, 2005; Viana et al., 2013b; Viana et al., 2013a). In the marine environment, biotic dispersal by herbivores (sea turtles, ducks and fish) has been reported for seagrasses (Sumoski and Orth, 2012) and passive transport of other organisms is also known to occur (e.g. sessile invertebrates transported by sea turtles and crabs (Winston, 2012)). The opposite process may also take place – e.g. anecdotal evidence suggests that green turtle hatchlings use kelp rafts for passive dispersal (Carr and Meylan, 1980).

In terrestrial systems, animal-mediated dispersal is often facilitated by rewards encasing the propagules or attached to the seeds (such as fruits, elaiosomes and some pods). In contrast, aquatic plant seeds typically lack rewards; hence, their dispersal is mediated by the ingestion of plant vegetative parts by herbivores ('foliage in the fruit', sensu Janzen (1984)), the occasional survival of seeds ingested by granivores (e.g. teals (Brochet et al., 2010b)) or the accidental ingestion of seeds by carnivores/omnivores (notably filter-feeders, such as flamingoes and shovelers (Verhoeven, 1980; Figuerola et al., 2003)). This difference results in major differences in selection pressures, which can be traced to differences in predominant seed traits (Table 1). Aquatic plant seeds are typically small, which facilitates their ingestion mixed with the foliage, their survival to gut passage, and a prolonged

360 gut-passage times resulting in longer dispersal distances (Mueller and van der Valk, 2002;
361 Charalambidou and Santamaria, 2005; Soons et al., 2008; Figuerola et al., 2010). Larger seeds often
362 have thick and impermeable coats, necessary to withstand their severe scarification in waterbird guts
363 (which tend to retain selectively such larger seeds (Kleyheeg, 2015) and the severe physico-chemical
364 treatment exerted by their guts (Figuerola et al., 2002; Santamaria et al., 2002). Such coat types
365 often result in strong physical dormancies, which may postpone seed germination until ingestion by a
366 potential dispersal vector has taken place (e.g. in seeds collected from the sediment bank by
367 granivores and filter feeders (Figuerola et al., 2003). Invertebrate propagules dispersed by waterfowl
368 typically show similar traits: small size, one to several protective coats, and delayed or stochastic
369 hatching (Charalambidou and Santamaria, 2002). In some cases, the encasement of resting eggs in
370 the mother's body may provide both protection against their digestion and a reward to potential
371 dispersers – having thus a function analogous to that of plant fruits, whose functional and adaptive
372 value of such a trait remains to be studied.

373 External dispersal may take place attached to the animal's fur or plumage, or adhered to mud-
374 stained surfaces in the body or feet (Figuerola and Green, 2002b; Frisch et al., 2007). Available
375 evidence suggests, however, that it is less frequent than internal dispersal (Brochet et al., 2010b).
376 Once again, small propagules are much more likely to become attached or adhered, and remain in
377 such situation long enough for long-distance dispersal to occur. Other traits that have been generally
378 assumed to facilitate external dispersal, such as flat shapes and suitable surface structures (hooks,
379 thorns, hairs), are known to occur in plant and invertebrate propagules (e.g. Vivian-Smith and Stiles
380 (1994)). As above, their functional and adaptive value remains to be studied.

381 Propagule dispersal, particularly over long distances, influences the population, genetic and
382 community structure of aquatic organisms (Viana et al., 2014). Hence, it is broadly regarded as a key
383 ecosystem service provided by aquatic herbivores. Its effect is, however, more evident when such
384 immigrants encounter empty niches (e.g. colonization of disturbed, new or restored wetlands) than

when they face resident genotypes or species, established before their arrival (Louette and De Meester, 2005). In such cases, the immigrant's establishment may be precluded by biotic resistance arising from intra- and inter-specific competition, mortality caused by natural enemies and environmental filtering (e.g. habitat changes caused by other species). Even then, the process of propagule dispersal represents a key element of ecosystem resilience – particularly in inland waters, which are fragmented and isolated by nature. Such resilience may prove vital, in the near future, for ecosystem adaptation to global change – by facilitating rapid range shifts and the readjustment of genetic structure (e.g. locally-adapted genotypes) caused by the accelerating changes in environmental conditions associated to global warming, land use changes and the perturbation of global nutrient cycles (Amezaga et al., 2002; Röckstrom et al., 2009; Robledo-Arnuncio et al., 2014).

Propagule dispersal by aquatic herbivores may also entail negative effects for native species and ecosystems when it mediates the arrival and spread of alien species (Reynolds et al., 2015). Examples include both plant and invertebrate species (Charalambidou et al., 2003; Brochet et al., 2009; Munoz et al., 2013), although the effect of dispersal on human-mediated dispersal is often predominant or difficult to disentangle (Weisz and Yan, 2010; Van Leeuwen et al., 2013). It is also worth noting that, despite their potential role as seed dispersers, aquatic herbivores may also reduce the frequency of propagule dispersal by reducing propagule production – due to the consumption of plant vegetative parts (e.g. (Wood et al., 2012a; Darnell and Dunton, 2015)) and the invertebrates attached or associated to them.

3.2.2 Biogeochemical cycling

Grazing and bioturbation by aquatic herbivores can have direct and indirect effects on biogeochemical cycling. In oligotrophic systems, grazing by smaller herbivores can have positive effects by conserving nutrients within the meadow and closing the cycling of nutrients from leaf material. Leaf material can be shredded (sea urchins, (Vonk et al., 2008b)), burrowed (Alpheid

410 shrimp, (Vonk et al., 2008a)) or excreted after grazing (fish, (Kirsch et al., 2002)), thus stimulating
411 nutrient retention. Larger herbivores such as green turtles and dugongs may travel between different
412 habitats and stimulate export of nutrients between foraging (seagrass) and resting (coral reefs) areas
413 (Christianen et al., 2012).

414 By foraging and resting in terrestrial and aquatic environments, herbivores provide aquatic-terrestrial
415 linkages, transporting carbon, nutrients and contaminants from land to water (for instance
416 hippopotamus *Hippopotamus amphibius* (Subalusky et al., 2015), waterbirds (Hahn et al., 2008;
417 Chaichana et al., 2010) or large savanna herbivores (Moss, 2015) or *vice versa* (for instance by moose
418 *Alces sp.* (Bump et al., 2009)). It has recently been suggested that the decline in large herbivore
419 densities and the extinction of Late Pleistocene megafauna caused a strong reduction in the capacity
420 of transport of phosphorus from nutrient hotspots, such as streams or floodplains, towards less
421 fertile inland areas (Doughty et al., 2016). Long distance travel of migratory herbivores also
422 contributes to transport of nutrients across sites of varying fertility (Bauer and Hoyer, 2014).

423 In seagrass meadows, grazing by mesoherbivores can increase productivity and possibly carbon
424 sequestration. However below-ground grazing (e.g. by dense populations of green turtles) or other
425 factors of disturbance in the seagrass root mat can cause release of ancient carbon, which may
426 contribute to increased global warming (Macreadie et al., 2015). Similarly, early season below-
427 ground foraging by pink-footed geese (*Anser brachyrhynchus*) is sufficient to strongly reduce C sink
428 strength and soil C stocks of arctic tundra (van der Wal et al., 2007). Recent studies are therefore
429 stressing that it is critical to maintain intact predator populations that control large herbivore
430 densities to prevent grazer aggregation, protect carbon stocks and avoid seagrass meadow collapse
431 (Atwood et al., 2015). In a subtropical seagrass ecosystem, large predators (e.g. tiger sharks) induce
432 plant species shifts by changing the foraging tactics of large grazers, such as turtle and dugongs
433 (Heithaus et al., 2007). Under low predation risk, dugongs and sea turtles foraged by excavating

nutrient-rich rhizomes of seagrasses. Under high predation risks, they changed their foraging tactics, which stimulated slow-growing pioneer species and enhanced carbon stocks.

Aquatic herbivores can also enhance methane emission through the damage of emergent plant stems (greylag geese (*Anser anser*) (Dingemans et al., 2011); grasshoppers (Petruzzella et al., 2015)). The stems of emergent macrophytes have well-developed lacunar systems for gas transport; hence, broken stems may act like “chimneys”, providing an open connection between the sediment and the atmosphere that bypasses the water layer. On the other hand, herbivores can reduce methane emission through bioturbation and removal of submerged plant beds (Bodelier et al., 2006).

3.2.3 Coastal protection

Seagrasses, mangroves and saltmarshes offer important coastal protection and sediment stabilization services. For seagrasses this function is generally attributed to seagrass canopy properties (Hendriks et al., 2010) and could be altered by herbivory. Although intensively grazed seagrass meadows have been shown to maintain their capacity for effective sediment stabilization, this function degrades when herbivores switch to belowground grazing, which causes decreased bed elevation, erosion and reduced coastal protection (Christianen et al., 2013). Similarly, livestock grazing can lower saltmarshes’ accretion rates (Nolte et al., 2015), weakening the resilience and coastal protection function of these systems.

3.2.4 Habitat for other organisms

Macrophytes play an important role in structuring aquatic communities because they provide physical structure, increase habitat complexity and heterogeneity, affect oxygen and nutrient concentrations and provide refuge from predation (Carpenter and Lodge, 1986; Jeppesen et al.,

1998). Macrophytes also release dissolved organic carbon (DOC) which can be used by microbes in the periphyton or plankton (Findlay et al., 1986). Therefore, by consuming macrophytes, herbivores may deteriorate the habitat for other organisms. Fish productivity was found to be lower in seagrass meadows grazed by green turtles (Arthur et al., 2013) and after meadow collapse caused by overgrazing (reported for turtles in (Christianen et al., 2013). Similarly, herbivorous fishes, dugongs, geese and other waterbirds have been found to drastically reduce invertebrate biomass in seagrass meadows and salt marshes (Marklund et al., 2002; Sherfy and Kirkpatrick, 2003; Skilleter et al., 2007; Pages et al., 2012) . Impacts on invertebrates may occur even where the proportional reduction of vegetation is low (Bortolus et al., 1998).

3.2.5. Primary production

Submerged and emergent macrophytes can significantly contribute to the primary production of aquatic ecosystems (Blindow et al., 2006; Brothers et al., 2013). This holds especially for small lakes which represent approximately 99% of all lakes (Downing et al., 2006; Verpoorter et al., 2014). Direct studies on the effect of herbivory on aquatic plant growth and production are scarce because quantification of the impact of grazing rates on plant production requires coupled measurements of age-dependent grazing loss and turnover rate of plant tissue (Sand-Jensen et al., 1994). In general, fast turnover of the plant tissue (i.e. high specific growth rate) can compensate for intense herbivory under non-limiting resource conditions (Sand-Jensen and Jacobsen, 2002). Cherry & Gough (2009) found that *Nymphaea odorata* may tolerate moderate levels of herbivory by reallocating biomass and resources aboveground. On the contrary, stands of *Myriophyllum spicatum* retract their resources to belowground parts after defoliation by aquatic caterpillars (Miler and Straile, 2010). Water hyacinths (*Eichhornia crassipes*) were also found to fully compensate for low levels of continuous defoliation, regardless of nutrient availability (Soti and Volin, 2010). In seagrass ecosystems, reported effects of large herbivores were positive for intermediate densities of green

turtles (e.g. an increased tolerance to eutrophication), and negative for high densities of green turtles (e.g. switch to belowground grazing causing meadow collapse) (Christianen et al., 2012; Christianen et al., 2013). Similarly, moderate levels of simulated fish herbivory stimulated seagrass in primary production (i.e. compensatory growth), while very high levels of herbivory decreased it (Verges et al., 2008).

At whole-ecosystem level, herbivory on submerged macrophytes may have different effects on gross primary production (GPP). If herbivory results in a shift from clear-water to turbid conditions (see below), GPP can be expected to decline, at least under intermediate concentrations of total phosphorus (Brothers et al., 2013). On the other hand, large grazers that remove old seagrass leaves covered in ephyphytes and have been reported to increase primary production (Valentine et al., 1997; Moran and Bjorndal, 2005; Christianen et al., 2012).

3.2.6. *Regime shifts*

Shallow eutrophic lakes and lowland rivers may exist in two alternative stable steady states, a clear-water state dominated by submerged macrophytes and a turbid, phytoplankton-dominated state (Scheffer et al., 1993; Hilt et al., 2011; Hilt, 2015). In deeper lakes, submerged macrophytes may also contribute to the stabilisation of clear-water conditions (Hilt et al., 2010; Sachse et al., 2014). Shifts between clear-water and turbid states have been attributed to changes in nutrient loading, in the abundance of zooplanktivorous fish (e.g. by biomanipulation) and/or in macrophyte cover (Scheffer et al., 1993; Sondergaard et al., 2007; Bakker et al., 2010). Herbivory on macrophytes may also play a significant role for shifting macrophyte-dominated systems into the turbid state, or preventing the shift from turbid- into clear-water conditions. Mitchell & Wass (1996a) concluded that the cumulative effect of waterfowl grazing consumption was small but might become critical when other conditions for macrophyte growth become limiting (due e.g. to light limitation caused by high water turbidity). A recent modeling study indeed showed that herbivory on macrophytes often becomes

important in combination with additional stress by periphyton shading (Hidding et al., 2016).
Herbivory by birds and fish may thus trigger the loss of submerged vegetation under high nutrient
loading (Van Donk and Otte, 1996; Paice et al., in press), possibly in combination with other stress
factors. Similarly, after reductions in nutrient loading, herbivorous birds may inhibit the expected
recovery of macrophytes (Lauridsen et al., 1993; Søndergaard et al., 1996; Hilt, 2006).

4. Perspectives: historical and future changes in herbivore grazing pressure

Herbivore management and global environmental change, including water level fluctuations,
eutrophication, temperature rise and invasive species, feedback on herbivore numbers, herbivore
distribution and grazing pressure.

4.1 Changes in herbivore assemblages over time

There is a growing body of evidence that herbivore assemblages have varied over time considerably
in their diversity and abundance, and are likely to continue to vary in the future. Throughout human
history, people have exploited many aquatic herbivore species, including waterfowl, Sirenians,
beavers, and muskrats, for food, recreation, and animal products such as skins, furs, feathers, and
oils (Domning, 1982; Kitchener and Conroy, 1997). Human overexploitation has had catastrophic
effects on many herbivore populations, with a wide range of species experiencing reduced
population sizes and geographic ranges, and even extinction (Jessen, 1970; Turvey and Risley, 2006).
Profound historical changes in herbivore assemblages were particularly evident in shallow seas and
coastal habitats, where the diversity and distributions of mammalian mega-herbivores (Sirenians
such as manatees and dugongs) were reduced heavily due to hunting by humans (Whitehead, 1978;
Jackson, 1997; Turvey and Risley, 2006; McCauley et al., 2015). Freshwater systems have also seen

the loss of many large-bodied herbivore species, in particular mammals (Moss, 2015; Bakker et al., 2016b). Additionally, during the twentieth century a range of semi-aquatic herbivore species switched from aquatic to terrestrial feeding, particularly during winter (Laubek, 1995), further reducing herbivore abundance and diversity within aquatic systems. Such habitat shifts have been most widely documented for avian herbivores such as species of swans, geese, ducks and rails (Jefferies et al., 2003; Van Eerden et al., 2005). The substantial effects of humans on aquatic herbivores meant that the twentieth century represented a low point for herbivore abundance and diversity across aquatic systems. Indeed, we find it interesting to note that earlier authors drew their conclusions on the apparent unimportance of plant-herbivore interactions in aquatic ecosystems based on research conducted during a period in which aquatic herbivores were relatively scarce. For Caribbean coastal ecosystems it has even been documented that these were severely degraded long before ecologists began to study them, through the decimation of large vertebrates including green turtles and manatees by about the year 1800 (Jackson 1997).

Stricter hunting regulations and conservation efforts in the second half of the twentieth century have facilitated recoveries in the range and population sizes of many key aquatic herbivore species (Nolet and Rosell, 1998). An example is the Eurasian beaver (*Castor fiber*), reduced by overhunting at the beginning of the twentieth century to c.1200 individuals in eight isolated populations across Europe (Nolet and Rosell, 1998; Halley and Rosell, 2002). Following greater legal protections from hunting, the Eurasian beaver underwent sustained population recovery and has re-established populations in all areas within its former natural range (with the exception of Portugal, Italy, and the southern Balkans), with a total population of at least 1.04 million individuals (Halley et al., 2012). Many species of herbivorous waterfowl in temperate regions have made similar recoveries (Bellrose, 1976; Ankney, 1996). Of the 21 goose species (*Anser* spp. and *Branta* spp.) for whose long-term population trends in Europe are known, 16 species are currently increasing (Fox et al., 2010). Recent changes in agricultural practices have resulted in greater terrestrial food availability for overwintering waterfowl, with larger species benefiting more in terms of population growth (Jefferies et al., 2003;

557 Van Eerden et al., 2005). Although many reptilian herbivores remain endangered, certain species
558 such as the green turtle (*Chelonia mydas*) have recently shown signs of population recovery in
559 response to three decades of conservation efforts (Chaloupka et al., 2008).

560 The recovery of species of aquatic herbivores has been aided by the recent interest in rewilding
561 ecosystems (Donlan et al., 2006). The key roles that extinct or extirpated large herbivores played in
562 the structure and functioning of terrestrial ecosystems has received growing recognition from
563 researchers (Donlan et al., 2006; Sandom et al., 2014; Bakker et al., 2016a; Doughty et al., 2016).
564 Recently, it has been proposed that large herbivores may have played equally-important roles in
565 regulating the structure and functioning of aquatic ecosystems (Moss, 2015; Bakker et al., 2016b).
566 Species of aquatic herbivore which can act as ecosystem engineers, such as the beaver, are typically
567 prime candidates for rewilding projects due to the wider ecosystem benefits that result from such
568 engineering (Collen and Gibson, 2001). The recent finding that beaver's creation of ponds increased
569 the diversity of herbivorous waterfowl within the landscape indicates that natural recolonization and
570 rewilding may result in wider changes to herbivore assemblages than the target species alone,
571 through the facilitation of different herbivore taxa (Nummi and Holopainen, 2014).

572 The recovery of predator populations, via natural recovery and conservation efforts, can also affect
573 plant-herbivore interactions (Estes et al., 2011). Evidence from terrestrial and marine systems shows
574 that herbivore impacts on plants can be reduced as predator numbers recover, because predators
575 not only lower herbivore abundance through direct consumption, but also alter herbivore
576 distributions and reduce grazing intensity through indirect effects of predator avoidance behavior
577 ('landscape of fear' *sensu* (Madin et al., 2011; Kuijper et al., 2013). Similar results could be found for
578 interactions between predators, herbivores, and macrophytes in aquatic systems, but little research
579 has been carried out to date. For example, a recent study found that green turtle (*Chelonia mydas*)
580 habitat use reflected trade-offs between food resources, body condition, and risk of predation by
581 tiger sharks (*Galeocerdo cuvier*) in seagrass beds (Heithaus et al., 2007). A decline in tiger shark

numbers may thus result in a strong increase of green turtle grazing on seagrass beds, potentially resulting in a seagrass bed collapse (Heithaus et al., 2014).

4.2 Exotic herbivore species

The spread of non-native species has been a key driver of temporal changes in aquatic herbivore assemblages in recent decades. A wide range of herbivore taxa have established invasive populations known to impact on native macrophytes. Well-documented examples include birds such as the mute swan in North America (Tatu et al., 2007), mammals such as the muskrat in Europe (Danell, 1979; Sarneel et al., 2014), fishes such as the lessepsian rabbitfishes in the Mediterranean (Verges et al., 2014a), molluscs such as the golden apple snail in Asia (Carlsson et al., 2004), and crustaceans such as red swamp crayfish in Europe (Gherardi and Acquistapace, 2007; Van der Wal et al., 2013). Despite attempts to prevent species transfer and establishment through improved biosecurity, rates of invasion remain at a historic high (Cohen and Carlton, 1998; Jackson and Grey, 2013).

4.3 Climate change and temperature rise

Future environmental change is also predicted to alter herbivore assemblages. In particular, climate-driven factors such as sea-level rise and changes in vegetation phenology and abundance have the potential to alter where, when and how much herbivores feed, and thus have the potential to alter plant-herbivore interactions across aquatic systems (Stillman et al., 2015). For example, northward shifts in wintering range in response to warming have been recorded for several waterfowl species, including semi-aquatic herbivores like greylag geese (Ramo et al., 2015). Furthermore, food requirements change with temperature. With increasing temperatures, the energy requirements of temperate ectotherm animals increase and they consume more food. Furthermore, they may change their diet in response to temperature, which has consequences for the degree of plant consumption.

Omnivorous fish increase their relative consumption of plant material with increasing temperatures (Prejs, 1984; Behrens and Lafferty, 2007, 2012), a pattern that is recently also found in copepods feeding on seston (Boersma et al., 2016). For endotherms this pattern may be the opposite, as their energy requirements decrease with increasing temperatures, but this hypothesis awaits empirical testing. Warmer temperatures may also induce higher periphyton shading of macrophytes (Mahdy et al., 2015), making them more prone to herbivory (Hidding et al., 2016).

4.4 Herbivore impacts under human control

Where herbivore impacts on macrophytes affect human activities, such as conservation, recreation, and aquaculture, they may be viewed as undesirable. In most cases it should be noted that problems caused by herbivores are often the direct or indirect consequence of earlier human actions. For instance, ongoing eutrophication of lakes reduces the resilience of submerged plant beds to grazing due to increased shading by periphyton which profits from the nutrient loading (Hidding et al., 2016), and re-establishment of submerged vegetation can be inhibited by grazers as well (Hauxwell et al., 2004). The combined hunting of large predators and the creation of marine protected reserves, as the place where large herbivores are safe, has locally resulted in very strong grazing pressure on seagrass beds by green sea turtles (Christianen et al. 2014).

Such herbivore impacts can lead to conflicts between people interested in herbivore welfare and conservation, and those interested in the activity being affected by the herbivore (Redpath et al., 2015). To date, conflicts have arisen due to overgrazing by reptilian, mammalian, and avian herbivores (Table 2). In contrast, we could find no evidence of conflicts associated with invertebrate herbivory, which may reflect differences in human values rather than ecological impact. Indeed, evidence from a recent meta-analysis showed that vertebrate herbivores do not have consistently greater impacts on macrophytes than invertebrate herbivores (Wood et al., 2016). Vertebrate herbivores such as waterfowl, turtles, and Sirenians are often considered charismatic, and attempts

to manage their numbers, behaviour or distributions are likely to attract more opposition from conservation and welfare groups than the management of invertebrate herbivores (Bremner and Park, 2007; Small, 2012). Consequently, it can be difficult to implement management to alleviate the effects of overgrazing (Coluccy et al., 2001).

Despite these conflicts, not all herbivore impacts on macrophytes are viewed as negative by people. Herbivores have proven to be effective biocontrol agents to help reduce and eradicate undesirable macrophytes, such as overabundant or invasive species (Newman, 2004; Cuda et al., 2008). A wide range of herbivore biocontrol agents have been used globally (Table 3). Biocontrol of macrophytes through herbivory has received growing interest from researchers and managers, particularly as some biotypes of invasive macrophytes (e.g. *Hydrilla verticillata*) have developed resistance to commonly-used herbicides (Cuda et al., 2008). More targeted biocontrol can be achieved by invertebrate herbivores, which typically show greater specificity for macrophyte species relative to vertebrate herbivores (Lodge, 1991; Newman, 1991). Since Wilson (1964) argued that “no insects have yet been used for the biological control of aquatic weeds”, a wide range of species of coleopteran, lepidopteran, and dipteran biocontrol agents have been used successfully (Newman, 2004; Cuda et al., 2008). Both the conflicts related to overgrazing, and the use of herbivores as biocontrol agents, show the importance of improving our understanding plant-herbivore interactions. For example, understanding the density-dependence of herbivore impacts on macrophyte abundance can inform the densities of biocontrol agents required to reduce overabundant macrophytes (Cuda et al., 2008), or allow lake managers to predict the response of macrophytes to changes in wild herbivore density (Wood et al., 2012a).

5. How to improve understanding of herbivore impacts

5.1 Bottom-up versus top-down control across environmental gradients

Lodge (1991) ended his review with the conclusion that “To understand the influence of herbivory (relative to other biotic and abiotic factors) on macrophyte populations and assemblages, extensive comparisons of grazing damage across environmental gradients and across macrophyte and grazer species must be made.” Whereas meta-analyses have provided valuable insight in the impact of different herbivores both on Angiosperms and macro-algae (Poore et al., 2012; Wood et al., 2016), and the relationship between macrophyte palatability and herbivore impact received increasing attention during the last decade, the variation in the intensity and impacts of herbivory along environmental gradients has been scantily explored. Bakker and Nolet (2014) suggested that herbivore impact may increase at nutrient-rich conditions, due to a combination of higher plant palatability and lower tolerance to grazing damage caused by additional stress factors (such as reduced light availability due to periphyton shading (Hidding et al., 2016). In an experimental pond system, herbivory by mallards proved to have stronger impact on submerged macrophytes under eutrophic than under oligotrophic conditions. We are not aware, however, of any field tests for this theory. Alternatively, herbivores may facilitate submerged macrophytes with increasing eutrophication, for example when moderate densities of small grazers such as freshwater snails clean submerged plants from epiphytes, which becomes more important under eutrophic conditions (Bakker et al., 2013). Also, when intense grazing stimulates the formation of new shoots, which are not yet colonized by epiphytes, large grazers such as green sea turtles, can compensate the negative effect of eutrophication for seagrass growth to a certain extent (Christianen et al., 2012).

5.2 Integrating marine and freshwater studies

Traditionally, freshwater macrophytes and seagrasses have largely been studied separately. To date, the literature on freshwater and marine herbivore impacts have not been integrated, and have largely developed separately despite the obvious areas of overlap. If cross-system comparisons were being made, these were often marine-terrestrial or freshwater-terrestrial comparisons and can

include both vascular plants and algae (Hay, 1991; Elser et al., 2000; Burkepile, 2013). This is unfortunate because, leaving a few obvious differences apart (such as the higher salinity and connectivity of marine systems), the ecology of marine and inland-water vascular macrophytes shows very few differences. Yet, integrative work on freshwater and marine vascular macrophytes still awaits its moment.

5.2 Herbivore assemblages: towards functional groups

For a better understanding of herbivore impact, researchers should consider the whole community of herbivores, which compete and facilitate each other. This has already been done for terrestrial systems long ago (McNaughton, 1985), but examples for aquatic systems are rare. One such example is the herbivore community in a shallow freshwater lake that forages on pondweed beds, consisting mainly of *Potamogeton pectinatus*. The sprouting plants are being grazed by resident waterfowl species like mute swans (*Cygnus olor*), mallards (*Anas platyrhynchos*), gadwall (*Anas strepera*) and coots (*Fulica atra*). The intensity and timing of grazing determine how much above-ground plant material is remaining and growing (Hootsmans, 1999). At the end of the summer, below-ground tubers are being formed depending on the amount of above-ground plant material (Van Wijk, 1988). Thus, more and, notably, earlier waterfowl grazing in summer results in less tuber biomass in autumn (Klaassen et al., 2006; Gyimesi et al., 2011). In autumn, tubers are being depleted by migratory swans (Bewick's swans *Cygnus columbianus*) and diving ducks (mainly tufted ducks *Aythya fuligula* and pochards *Aythya ferina*) until a grazing threshold (Nolet et al., 2001). The diving ducks benefit from the trampling activity of the swans, without negatively affecting the swans' intake rate; this therefore classifies as commensalism (Gyimesi et al., 2012). In accordance with a sequential population model (Jonzén et al., 2002), highest tuber biomass and in particular tuber production was generally found at sites foraged down to intermediate thresholds in the previous autumn. In this system a positive

feedback between tuber grazing and tuber production resulted from a reduction in self-shading or a decrease in neighbour competition (Nolet, 2004).

Together, the interactions within herbivore communities will determine the effect of herbivore diversity, a topic rarely touched upon in aquatic vascular plant beds (but see marine macro-algae and seagrass systems, e.g (Duffy et al., 2003; Burkepile, 2013). Whereas all interactions at the species level are interesting, a way forward can be to generalize beyond species by grouping herbivores in guilds or functional groups and working out which traits best explain their relative effects. Such traits could include body size, diet (herbivory-omnivory), habitat (terrestrial-aquatic linkages), migratory strategy (sedentary versus migratory) and movement ecology (foraging ranges). Recently, grouping of large savanna herbivores proved to be useful to understand their ecosystem impacts (Hempson et al., 2015). Similarly, the ecosystem functions of aquatic large herbivores may be understood from their habitat use, in particular how dependent on the aquatic systems they are in combination with their movement ecology (Bakker et al., 2016b).

5.3 Tools to study herbivore impacts

Practically, the high variance in estimates of macrophyte abundance, even within a single study system, can make herbivore effects difficult to detect in aquatic systems without large sample sizes (Wood et al., 2012b). However, numerous methods have been developed and employed to detect and/or quantify herbivory on macrophytes and seagrasses (Table 4). The most direct approach for detection and quantification of effects is a technique known as tethering (Kirsch et al., 2002; Tomas et al., 2005; Prado et al., 2007; Pages et al., 2014). With this technique it is possible to estimate the biomass (or cm of leaf) eaten daily, i.e. direct herbivory rates (see Table 4). Another direct approach to detect and quantify herbivore effects are in situ enclosure cages with subsequent biomass measurements in- and outside.

Other less direct herbivory detection and quantification methods focusing on macrophytes include visual estimations of leaf damage and measurements of macrophyte performance across naturally-occurring spatial or temporal gradients in herbivore assemblage properties (e.g. density) (Table 4). Aerial photographs taken from drones (Brandt et al., 2015), echo-sounding (Jäger et al., 2004) and remote sensing (Silva et al., 2008) may increasingly being used in the future for larger scale quantifications of macrophyte consumption by herbivores. At the herbivore side, underwater videos, aquarium feeding experiments, molecular markers, stable isotope signatures, gut and faeces analyses, and whole-lake fish telemetry have been applied (Table 4).

Finally, mechanistic models allow assessments of herbivory over larger spatial and temporal scales than field-based methods, e.g. long-term predictions of future herbivory. They initially need to be tested against field data to demonstrate accuracy and have been developed for both, the herbivores and the plants (Table 4). Recently, a plant growth models for a specific macrophyte species has been used to detect a synergy between herbivory and shading by periphyton as additional stressor (Hidding et al., 2016). Simulation models of herbivore foraging can be useful tools to predict foraging impacts, and test strategies for grazing management (Wood et al., 2014a; Nolet et al., 2016).

6. Conclusions

Over the last 25 years, a substantial body of evidence has developed that shows that herbivory is an important factor in the ecology of macrophytes across freshwater and marine habitats. Compiling the most recent data, we conclude that herbivore impacts in freshwater and marine ecosystems are typically 5-10 times greater than those reported for terrestrial ecosystems. This corresponds with lower C:N stoichiometry of submerged aquatic plants. Furthermore, aquatic habitats are characterized by large variation in grazing pressure. Considerable changes have occurred, and are predicted to occur, in herbivore diversity and abundance, with wide implications for the composition and dynamics of macrophyte communities, as well as for the structure and functioning of aquatic

ecosystems. There are pressing needs to improve our management of undesirable herbivore impacts on macrophytes (e.g. leading to an ecosystem collapse), and the conflicts between people associated with the impacts of charismatic mega-herbivores. While simultaneously, the long-term future of maintaining both viable herbivore populations and plant beds should be addressed, as both belong in complete ecosystems and have co-evolved in these long before the increasing influence of man.

Most research to date has focused on the short-term impacts of herbivores on macrophyte abundance and community composition. To understand the roles of herbivores more fully we need to consider their longer-term impacts and the role of herbivory in the (co-)evolution of both macrophyte and herbivore species. Furthermore, a better integration of the freshwater, marine, and terrestrial herbivory literatures would greatly benefit future research efforts.

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Literature

- Allsopp, W.H.L., 1960. The manatee: ecology and use for weed control. *Nature* 188, 762.
- Amezaga, J.M., Santamaria, L., Green, A.J., 2002. Biotic wetland connectivity - supporting a new approach for wetland policy. *Acta Oecol.* 23, 213-222.
- Ankney, C.D., 1996. An embarrassment of riches: Too many geese. *J. Wildlife Manage.* 60, 217-223.
- Aragones, L.V., Lawler, I.R., Foley, W.J., Marsh, H., 2006. Dugong grazing and turtle cropping: grazing optimization in tropical seagrass systems? *Oecologia* 149, 635-647.
- Arthur, R., Kelkar, N., Alcoverro, T., Madhusudan, M.D., 2013. Complex ecological pathways underlie perceptions of conflict between green turtles and fishers in the Lakshadweep Islands. *Biol. Conserv.* 167, 25-34.
- Atkinson, M.J., Smith, S.V., 1983. C-N-P ratios of benthic marine plants. *Limnol. Oceanogr.* 28, 568-574.
- Atwood, T.B., Connolly, R.M., Ritchie, E.G., Lovelock, C.E., Heithaus, M.R., Hays, G.C., Fourqurean, J.W., Macreadie, P.I., 2015. Predators help protect carbon stocks in blue carbon ecosystems. *Nature Climate Change* 5, 1038-1045.
- Bakker, E.S., Dobrescu, I., Straile, D., Holmgren, M., 2013. Testing the stress gradient hypothesis in herbivore communities: facilitation peaks at intermediate nutrient levels. *Ecology* 94, 1776-1784.
- Bakker, E.S., Gill, J.L., Johnson, C.N., Vera, F.W.M., Sandom, C.J., Asner, G.P., Svenning, J.C., 2016a. Combining paleo-data and modern exclosure experiments to assess the impact of megafauna extinctions on woody vegetation. *P. Natl. Acad. Sci. USA* 113, 847-855.
- Bakker, E.S., Nolet, B.A., 2014. Experimental evidence for enhanced top-down control of freshwater macrophytes with nutrient enrichment. *Oecologia* 176, 825-836.

800 Bakker, E.S., Pagès, J.F., Arthur, R., Alcoverro, T., 2016b. Assessing the role of large herbivores in the
801 structuring and functioning of freshwater and marine angiosperm ecosystems. *Ecography* 39,
802 162-179.

803 Bakker, E.S., Sarneel, J.M., Gulati, R.D., Liu, Z.W., van Donk, E., 2013. Restoring macrophyte diversity
804 in shallow temperate lakes: biotic versus abiotic constraints. *Hydrobiologia* 710, 23-37.

805 Bakker, E.S., Van Donk, E., Declerck, S.A.J., Helmsing, N.R., Hidding, B., Nolet, B.A., 2010. Effect of
806 macrophyte community composition and nutrient enrichment on plant biomass and algal
807 blooms. *Basic Appl. Ecol.* 11, 432-439.

808 Barends, F., 2002. The Muskrat (*Ondatra zibethicus*): expansion and control in the Netherlands. *Lutra*
809 45, 97-104.

810 Bauer, S., Hoyer, B.J., 2014. Migratory animals couple biodiversity and ecosystem functioning
811 Worldwide. *Science* 344, 54-+.

812 Bazely, D.R., Jefferies, R.L., 1985. Goose feces - a source of nitrogen for plant-growth in a grazed salt-
813 marsh. *J. Appl. Ecol.* 22, 693-703.

814 Behrens, M.D., Lafferty, K.D., 2007. Temperature and diet effects on omnivorous fish performance:
815 implications for the latitudinal diversity gradient in herbivorous fishes. *Can. J. Fish. Aquat. Sci.*
816 64, 867-873.

817 Behrens, M.D., Lafferty, K.D., 2012. Geographic Variation in the diet of Opaleye (*Girella nigricans*)
818 with respect to temperature and habitat. *PLoS One* 7, e45901.

819 Bellrose, F.C., 1976. The comeback of the wood duck. *Wildlife Soc. B.* 4, 107-110.

820 Bennett, S., Bellwood, D.R., 2011. Latitudinal variation in macroalgal consumption by fishes on the
821 Great Barrier Reef. *Mar. Ecol.-Prog. Ser.* 426, 241-U269.

822 Blindow, I., Hargeby, A., Meyercordt, J., Schubert, H., 2006. Primary production in two shallow lakes
823 with contrasting plant form dominance: A paradox of enrichment? *Limnol. Oceanogr.* 51,
824 2711-2721.

825 Bodelier, P.L.E., Stomp, M., Santamaria, L., Klaassen, M., Laanbroek, H.J., 2006. Animal-plant-microbe
826 interactions: direct and indirect effects of swan foraging behaviour modulate methane
827 cycling in temperate shallow wetlands. *Oecologia* 149, 233-244.

828 Boersma, M., Mathew, K.A., Niehoff, B., Schoo, K.L., Franco-Santos, R.M., Meunier, C.L., 2016.
829 Temperature driven changes in the diet preference of omnivorous copepods: no more meat
830 when it's hot? *Ecol. Lett.* 19, 45-53.

831 Borer, E.T., Seabloom, E.W., Shurin, J.B., Anderson, K.E., Blanchette, C.A., Broitman, B., Cooper, S.D.,
832 Halpern, B.S., 2005. What determines the strength of a trophic cascade? *Ecology* 86, 528-537.

833 Bortolus, A., Iribarne, O.O., Martinez, M.M., 1998. Relationship between waterfowl and the seagrass
834 *Ruppia maritima* in a southwestern Atlantic coastal lagoon. *Estuaries* 21, 710-717.

835 Bownes, A., 2014. Suitability of a leaf-mining fly, *Hydrellia* sp., for biological control of the invasive
836 aquatic weed, *Hydrilla verticillata* in South Africa. *Biocontrol* 59, 771-780.

837 Bownes, A., Hill, M.P., Byrne, M.J., 2010. Evaluating the impact of herbivory by a grasshopper,
838 *Cornops aquaticum* (Orthoptera: Acrididae), on the competitive performance and biomass
839 accumulation of water hyacinth, *Eichhornia crassipes* (Pontederiaceae). *Biol. Control* 53, 297-
840 303.

841 Brandt, E.C., Petersen, J.E., Grossman, J.J., Allen, G.A., Benzing, D.H., 2015. Relationships between
842 spatial metrics and plant diversity in constructed freshwater wetlands. *PLoS One* 10,
843 e0135263.

844 Bremner, A., Park, K., 2007. Public attitudes to the management of invasive non-native species in
845 Scotland. *Biol. Conserv.* 139, 306-314.

846 Brochet, A.L., Gauthier-Clerc, M., Guillemain, M., Fritz, H., Waterkeyn, A., Baltanas, A., Green, A.J.,
847 2010a. Field evidence of dispersal of branchiopods, ostracods and bryozoans by teal (*Anas*
848 *crecca*) in the Camargue (southern France). *Hydrobiologia* 637, 255-261.

849 Brochet, A.L., Guillemain, M., Fritz, H., Gauthier-Clerc, M., Green, A.J., 2009. The role of migratory
850 ducks in the long-distance dispersal of native plants and the spread of exotic plants in
851 Europe. *Ecography* 32, 919-928.

852 Brochet, A.L., Guillemain, M., Fritz, H., Gauthier-Clerc, M., Green, A.J., 2010b. Plant dispersal by teal
853 (*Anas crecca*) in the Camargue: duck guts are more important than their feet. *Freshw. Biol.*
854 55, 1262-1273.

855 Brothers, S.M., Hilt, S., Meyer, S., Kohler, J., 2013. Plant community structure determines primary
856 productivity in shallow, eutrophic lakes. *Freshw. Biol.* 58, 2264-2276.

857 Bump, J.K., Tischler, K.B., Schrank, A.J., Peterson, R.O., Vucetich, J.A., 2009. Large herbivores and
858 aquatic-terrestrial links in southern boreal forests. *J. Anim. Ecol.* 78, 338-345.

859 Burkepile, D.E., 2013. Comparing aquatic and terrestrial grazing ecosystems: is the grass really
860 greener? *Oikos* 122, 306-312.

861 Carlsson, N.O.L., Brönmark, C., Hansson, L.A., 2004. Invading herbivory: The golden apple snail alters
862 ecosystem functioning in Asian wetlands. *Ecology* 85, 1575-1580.

863 Carpenter, S.R., Lodge, D.M., 1986. Effects of submersed macrophytes on ecosystem processes.
864 *Aquat. Bot.* 26, 341-370

865 Carr, A., Meylan, A.B., 1980. Evidence of passive migration of green turtle hatchlings in *Sargassum*.
866 *Copeia*, 366-368.

867 Cebrian, J., Duarte, C.M., 1998. Patterns in leaf herbivory on seagrasses. *Aquat. Bot.* 60, 67-82.

868 Cebrian, J., Lartigue, J., 2004. Patterns of herbivory and decomposition in aquatic and terrestrial
869 ecosystems. *Ecol. Monogr.* 74, 237-259.

870 Cebrian, J., Shurin, J.B., Borer, E.T., Cardinale, B.J., Ngai, J.T., Smith, M.D., Fagan, W.F., 2009.
871 Producer nutritional quality controls ecosystem trophic structure. *PLoS One* 4, e4929.

872 Chaichana, R., Leah, R., Moss, B., 2010. Birds as eutrophication agents: a nutrient budget for a small
873 lake in a protected area. *Hydrobiologia* 646, 111-121.

874 Chaloupka, M., Bjorndal, K.A., Balazs, G.H., Bolten, A.B., Ehrhart, L.M., Limpus, C.J., Suganuma, H.,
 875 Troeeng, S., Yamaguchi, M., 2008. Encouraging outlook for recovery of a once severely
 876 exploited marine megaherbivore. *Global Ecol. Biogeogr.* 17, 297-304.

877 Charalambidou, I., Ketelaars, H.A.M., Santamaria, L., 2003. Endozoochory by ducks: influence of
 878 developmental stage of *Bythotrephes* diapause eggs on dispersal probability. *Divers. Distrib.*
 879 9, 367-374.

880 Charalambidou, I., Santamaria, L., 2002. Waterbirds as endozoochorous dispersers of aquatic
 881 organisms: a review of experimental evidence. *Acta Oecol.* 23, 165-176.

882 Charalambidou, I., Santamaria, L., 2005. Field evidence for the potential of waterbirds as dispersers
 883 of aquatic organisms. *Wetlands* 25, 252-258.

884 Cherry, J.A., Gough, L., 2009. Trade-offs in plant responses to herbivory influence trophic routes of
 885 production in a freshwater wetland. *Oecologia* 161, 549-557.

886 Christianen, M.J.A., Govers, L.L., Bouma, T.J., Kiswara, W., Roelofs, J.G.M., Lamers, L.P.M., van
 887 Katwijk, M.M., 2012. Marine megaherbivore grazing may increase seagrass tolerance to high
 888 nutrient loads. *J. Ecol.* 100, 546-560.

889 Christianen, M.J.A., Herman, P.M.J., Bouma, T.J., Lamers, L.P.M., van Katwijk, M.M., van der Heide,
 890 T., Mumby, P.J., Silliman, B.R., Engelhard, S.L., de Kerk, M.V., Kiswara, W., van de Koppel, J.,
 891 2014. Habitat collapse due to overgrazing threatens turtle conservation in marine protected
 892 areas. *P.R. Soc. B-Biol. Sci.* 281, 20132890.

893 Christianen, M.J.A., van Belzen, J., Herman, P.M.J., van Katwijk, M.M., Lamers, L.P.M., van Leent,
 894 P.J.M., Bouma, T.J., 2013. Low-canopy seagrass beds still provide important coastal
 895 protection services. *PLoS One* 8, e62413.

896 Clayton, J.S., 1996. Aquatic weeds and their control in New Zealand lakes. *Lake Reserv. Manage.* 12,
 897 477-486.

898 Clements, K.D., Raubenheimer, D., Choat, J.H., 2009. Nutritional ecology of marine herbivorous
 899 fishes: ten years on. *Funct. Ecol.* 23, 79-92.

900 Cloern, J.E., Canuel, E.A., Harris, D., 2002. Stable carbon and nitrogen isotope composition of aquatic
 901 and terrestrial plants of the San Francisco Bay estuarine system. *Limnol. Oceanogr.* 47, 713-
 902 729.

903 Coetzee, J.A., Byrne, M.J., Hill, M.P., 2007. Impact of nutrients and herbivory by *Eccritotarsus*
 904 *catarinensis* on the biological control of water hyacinth, *Eichhornia crassipes*. *Aquat. Bot.* 86,
 905 179-186.

906 Cohen, A.N., Carlton, J.T., 1998. Accelerating invasion rate in a highly invaded estuary. *Science* 279,
 907 555-558.

908 Collen, P., Gibson, R.J., 2001. The general ecology of beavers (*Castor* spp.), as related to their
 909 influence on stream ecosystems and riparian habitats, and the subsequent effects on fish - a
 910 review. *Rev. Fish Biol. Fisher.* 10, 439-461.

911 Coluccy, J.M., Drobney, R.D., Graber, D.A., Sheriff, S.L., Witter, D.J., 2001. Attitudes of central
 912 Missouri residents toward local giant Canada geese and management alternatives. *Wildlife*
 913 *Soc. Bull.* 29, 116-123.

914 Creed, R.P., Sheldon, S.P., 1993. The effect of feeding by a North American weevil, *Euhrychiopsis*
 915 *lecontei*, on Eurasian watermilfoil (*Myriophyllum spicatum*). *Aquat. Bot.* 45, 245-256.

916 Cronin, G., Lodge, D.M., Hay, M.E., Miller, M., Hill, A.M., Horvath, T., Bolser, R.C., Lindquist, N., Wahl,
 917 M., 2002. Crayfish feeding preferences for fresh water macrophytes: The influence of plant
 918 structure and chemistry. *J. Crust. Biol.* 22, 708-718.

919 Cuda, J.P., Charudattan, R., Grodowitz, M.J., Newman, R.M., Shearer, J.F., Tamayo, M.L., Villegas, B.,
 920 2008. Recent advances in biological control of submersed aquatic weeds. *J. Aquat. Plant*
 921 *Manage.* 46, 15-32.

922 Cyr, H., Pace, M.L., 1993. Magnitude and patterns of herbivory in aquatic and terrestrial ecosystems.
 923 *Nature* 361, 148-150.

924 Danell, K., 1979. Reduction of aquatic vegetation following the colonization of a Northern Swedish
 925 Lake by the muskrat, *Ondatra zibethica*. *Oecologia* 38, 101-106.

926 Darnell, K.M., Dunton, K.H., 2015. Consumption of turtle grass seeds and seedlings by crabs in the
 927 western Gulf of Mexico. *Mar. Ecol.-Prog. Ser.* 520, 153-163.

928 De Kluijver, A., Ning, J., Liu, Z., Jeppesen, E., Gulati, R.D., Middelburg, J.J., 2015. Macrophytes and
 929 periphyton carbon subsidies to bacterioplankton and zooplankton in a shallow eutrophic lake
 930 in tropical China. *Limnol. Oceanogr.* 60, 375-385.

931 Dingemans, B.J.J., Bakker, E.S., Bodelier, P.L.E., 2011. Aquatic herbivores facilitate the emission of
 932 methane from wetlands. *Ecology* 92, 1166-1173.

933 Domning, D.P., 1982. Commercial exploitation of manatees *Trichechus* in Brazil circa 1785–1973. *Biol.*
 934 *Conserv.* 22, 101-126.

935 Donlan, C.J., Berger, J., Bock, C.E., Bock, J.H., Burney, D.A., Estes, J.A., Foreman, D., Martin, P.S.,
 936 Roemer, G.W., Smith, F.A., Soule, M.E., Greene, H.W., 2006. Pleistocene rewilding: An
 937 optimistic agenda for twenty-first century conservation. *Am. Nat.* 168, 660-681.

938 Dorenbosch, M., Bakker, E.S., 2012. Effects of contrasting omnivorous fish on submerged
 939 macrophyte biomass in temperate lakes: a mesocosm experiment. *Freshw. Biol.* 57, 1360-
 940 1372.

941 Dos Santos, V.M., Matheson, F.E., Pilditch, C.A., Elger, A., 2012. Is black swan grazing a threat to
 942 seagrass? Indications from an observational study in New Zealand. *Aquat. Bot.* 100, 41-50.

943 Doughty, C.E., Roman, J., Faurby, S., Wolf, A., Haque, A., Bakker, E.S., Malhi, Y., Dunning Jr., J.B.,
 944 Svenning, J.C., 2016. Global nutrient transport in a world of giants. *P. Natl. Acad. Sci USA* 113,
 945 868-873.

946 Downing, J.A., Prairie, Y.T., Cole, J.J., Duarte, C.M., Tranvik, L.J., Striegl, R.G., McDowell, W.H.,
 947 Kortelainen, P., Caraco, N.F., Melack, J.M., Middelburg, J.J., 2006. The global abundance and
 948 size distribution of lakes, ponds, and impoundments. *Limnol. Oceanogr.* 51, 2388-2397.

949 Duarte, C.M., 1990. Seagrass nutrient content. *Mar. Ecol.-Prog. Ser.* 67, 201-207.

950 Duarte, C.M., Cebrian, J., 1996. The fate of marine autotrophic production. *Limnol. Oceanogr.* 41,
 951 1758-1766.

952 Duffy, J.E., Richardson, J.P., Canuel, E.A., 2003. Grazer diversity effects on ecosystem functioning in
 953 seagrass beds. *Ecol. Lett.* 6, 637-645.

954 Eigemann, F., Mischke, U., Hupfer, M., Hilt, S., 2016. Biological indicators track differential response
 955 of pelagic and littoral areas to nutrient load reduction in German lakes. *Ecol. Indic.* 61, 905-
 956 910.

957 Elser, J.J., Fagan, W.F., Denno, R.F., Dobberfuhl, D.R., Folarin, A., Huberty, A., Interlandi, S., Kilham,
 958 S.S., McCauley, E., Schulz, K.L., Siemann, E.H., Sterner, R.W., 2000. Nutritional constraints in
 959 terrestrial and fresh water food webs. *Nature* 408, 578-580.

960 Estes, J.A., Terborgh, J., Brashares, J.S., Power, M.E., Berger, J., Bond, W.J., Carpenter, S.R., Essington,
 961 T.E., Holt, R.D., Jackson, J.B.C., Marquis, R.J., Oksanen, L., Oksanen, T., Paine, R.T., Pickett,
 962 E.K., Ripple, W.J., Sandin, S.A., Scheffer, M., Schoener, T.W., Shurin, J.B., Sinclair, A.R.E.,
 963 Soule, M.E., Virtanen, R., Wardle, D.A., 2011. Trophic downgrading of planet earth. *Science*
 964 333, 301-306.

965 Figuerola, J., Charalambidou, I., Santamaria, L., Green, A.J., 2010. Internal dispersal of seeds by
 966 waterfowl: effect of seed size on gut passage time and germination patterns.
 967 *Naturwissenschaften* 97, 555-565.

968 Figuerola, J., Green, A.J., 2002a. Dispersal of aquatic organisms by waterbirds: a review of past
 969 research and priorities for future studies. *Freshw. Biol.* 47, 483-494.

970 Figuerola, J., Green, A.J., 2002b. How frequent is external transport of seeds and invertebrate eggs by
 971 waterbirds? A study in Donana, SW Spain. *Arch. Hydrobiol.* 155, 557-565.

972 Figuerola, J., Green, A.J., 2005. Effects of premigratory fasting on the potential for long distance
 973 dispersal of seeds by waterfowl: An experiment with marbled teal. *Revue D Ecologie-La Terre*
 974 *Et La Vie* 60, 283-287.

975 Figuerola, J., Green, A.J., Santamaria, L., 2002. Comparative dispersal effectiveness of wigeon grass
 976 seeds by waterfowl wintering in south-west Spain: quantitative and qualitative aspects. *J.*
 977 *Ecol.* 90, 989-1001.

978 Figuerola, J., Green, A.J., Santamaria, L., 2003. Passive internal transport of aquatic organisms by
 979 waterfowl in Donana, south-west Spain. *Glob. Ecol. Biogeogr.* 12, 427-436.

980 Findlay, S., Carlough, L., Crocker, M.T., Gill, H.K., Meyer, J.L., Smith, P.J., 1986. Bacterial growth on
 981 macrophyte leachate and fate of bacterial production. *Limnol. Oceanogr.* 31, 1335-1341.

982 Fourqurean, J.W., Jones, R.D., Zieman, J.C., 1993. Processes influencing water column nutrient
 983 characteristics and phosphorus limitation of phytoplankton biomass in Florida Bay FL, USA -
 984 inferences from spatial distributions. *Estuar. Coast. Shelf S.* 36, 295-314.

985 Fourqurean, J.W., Manuel, S., Coates, K.A., Kenworthy, W.J., Smith, S.R., 2010. Effects of excluding
 986 sea turtle herbivores from a seagrass bed: Overgrazing may have led to loss of seagrass
 987 meadows in Bermuda. *Mar. Ecol.-Prog. Ser.* 419, 223-232.

988 Fox, A.D., Ebbinge, B.S., Mitchell, C., Heinicke, T., Aarvark, T., Colhoun, K., Clausen, P., Dereliev, S.,
 989 Farago, S., Koffijberg, K., Kruckenberg, H., Loonen, M.J.J.E., Madsen, J., Mooij, J., Musil, P.,
 990 Nilsson, L., Pihl, S., van der Jeugd, H., 2010. Current estimates of goose population sizes in
 991 western Europe, a gap analysis and an assessment of trends. *Ornis Svecica* 20, 115-127.

992 France, R.L., 1996. Stable isotopic survey of the role of macrophytes in the carbon flow of aquatic
 993 foodwebs. *Vegetatio* 124, 67-72.

994 Franceschini, M.C., de Neiff, A.P., Galassi, M.E., 2010. Is the biomass of water hyacinth lost through
 995 herbivory in native areas important? *Aquat. Bot.* 92, 250-256.

996 Frank, D.A., Groffman, P.M., Evans, R.D., Tracy, B.F., 2000. Ungulate stimulation of nitrogen cycling
 997 and retention in Yellowstone Park grasslands. *Oecologia* 123, 116-121.

998 Frisch, D., Green, A.J., Figuerola, J., 2007. High dispersal capacity of a broad spectrum of aquatic
 999 invertebrates via waterbirds. *Aquat. Sci.* 69, 568-574.

1000 Gherardi, F., Acquistapace, P., 2007. Invasive crayfish in Europe: the impact of *Procambarus clarkii* on
 1001 the littoral community of a Mediterranean lake. *Freshw. Biol.* 52, 1249-1259.

1002 Gosling, L.M., Baker, S.J., 1989. The eradication of muskrats and coypus from Britain. *Biol. J. Linn. Soc.*
 1003 38, 39-51.

1004 Green, A.J., Figuerola, J., 2005. Recent advances in the study of long-distance dispersal of aquatic
 1005 invertebrates via birds. *Divers. Distrib.* 11, 149-156.

1006 Grey, J., Jackson, M.C., 2012. 'Leaves and eats shoots': direct terrestrial feeding can supplement
 1007 invasive red swamp crayfish in times of need. *PLoS One* 7, e42575.

1008 Gross, E.M., Bakker, E.S., 2012. The role of plant secondary metabolites in freshwater macrophyte-
 1009 herbivore interactions: limited or unexplored chemical defences? . In: Iason, G.R., Dicke, M.,
 1010 Hartley, S.E. (Eds.), *The Ecology of Plant Secondary Metabolites: From Genes to Global*
 1011 *Processes*. Cambridge University Press, British Ecological Society, pp. 154-169.

1012 Gross, E.M., Johnson, R.L., Hairston, N.G., 2001. Experimental evidence for changes in submersed
 1013 macrophyte species composition caused by the herbivore *Acentria ephemerella*
 1014 (Lepidoptera). *Oecologia* 127, 105-114.

1015 Gruner, D.S., Smith, J.E., Seabloom, E.W., Sandin, S.A., Ngai, J.T., Hillebrand, H., Harpole, W.S., Elser,
 1016 J.J., Cleland, E.E., Bracken, M.E.S., Borer, E.T., Bolker, B.M., 2008. A cross-system synthesis of
 1017 consumer and nutrient resource control on producer biomass. *Ecol. Lett.* 11, 740-755.

1018 Gyimesi, A., de Vries, P.P., de Boer, T., Nolet, B.A., 2011. Reduced tuber banks of fennel pondweed
 1019 due to summer grazing by waterfowl. *Aquat. Bot.* 94, 24-28.

1020 Gyimesi, A., van Lith, B., Nolet, B.A., 2012. Commensal foraging with Bewick's Swans *Cygnus bewickii*
 1021 doubles instantaneous intake rate of Common Pochards *Aythya ferina*. *Ardea* 100, 55-62.

1022 Hacker, S.D., Bertness, M.D., 1995. A herbivore paradox - why salt-marsh aphids live on poor-quality
 1023 plants. *Am. Nat.* 145, 192-210.

1024 Hahn, S., Bauer, S., Klaassen, M., 2008. Quantification of allochthonous nutrient input into
 1025 freshwater bodies by herbivorous waterbirds. *Freshw. Biol.* 53, 181-193.

1026 Halley, D., Rosell, F., Saveljev, A., 2012. Population and distribution of Eurasian Beaver (*Castor fiber*).
 1027 *Balt. For.* 18, 168-175.

1028 Halley, D.J., Rosell, F., 2002. The beaver's reconquest of Eurasia: status, population development and
 1029 management of a conservation success. *Mammal Rev.* 32, 153-178.

1030 Hangelbroek, H.H., Ouborg, N.J., Santamaria, L., Schwenk, K., 2002. Clonal diversity and structure
 1031 within a population of the pondweed *Potamogeton pectinatus* foraged by Bewick's swans.
 1032 Mol. Ecol. 11, 2137-2150.

1033 Hanlon, S.G., Hoyer, M.V., Cichra, C.E., Canfield, D.E., 2000. Evaluation of macrophyte control in 38
 1034 Florida lakes using triploid grass carp. J. Aquat. Plant Manage. 38, 48-54.

1035 Hanson, K.C., Cooke, S.J., Suski, C.D., Niezgoda, G., Phelan, F.J.S., Tinline, R., Philipp, D.P., 2007.
 1036 Assessment of largemouth bass (*Micropterus salmoides*) behaviour and activity at multiple
 1037 spatial and temporal scales utilizing a whole-lake telemetry array. Hydrobiologia 582, 243-
 1038 256.

1039 Hauxwell, J., Osenberg, C.W., Frazer, T.K., 2004. Conflicting management goals: Manatees and
 1040 invasive competitors inhibit restoration of a native macrophyte. Ecol. Appl. 14, 571-586.

1041 Havel, J.E., Bruckerhoff, L.A., Funkhouser, M.A., Gemberling, A.R., 2014. Resistance to desiccation in
 1042 aquatic invasive snails and implications for their overland dispersal. Hydrobiologia 741, 89-
 1043 100.

1044 Hay, M.E., 1991. Marine terrestrial contrasts in the ecology of plant-chemical defenses against
 1045 herbivores. Trends Ecol. Evol. 6, 362-365.

1046 Hay, M.E., 1996. Marine chemical ecology: What's known and what's next? J. Exp. Mar. Biol. Ecol.
 1047 200, 103-134.

1048 Heithaus, M.R., Alcoverro, T., Arthur, R., Burkholder, D.A., Coates, K.A., Christianen, M.J., Kelkar, N.,
 1049 Manuel, S.A., Wirsing, A.J., Kenworthy, W.J., Fourqurean J.W., 2014. Seagrasses in the age of
 1050 sea turtle conservation and shark overfishing. Frontiers in Marine Science 1, 28.

1051 Heithaus, M.R., Frid, A., Wirsing, A.J., Dill, L.M., Fourqurean, J.W., Burkholder, D., Thomson, J.,
 1052 Bejder, L., 2007. State-dependent risk-taking by green sea turtles mediates top-down effects
 1053 of tiger shark intimidation in a marine ecosystem. J. Anim. Ecol. 76, 837-844.

1054 Hempson, G.P., Archibald, S., Bond, W.J., 2015. A continent-wide assessment of the form and
 1055 intensity of large mammal herbivory in Africa. Science 350, 1056-1061.

1056 Hendriks, I.E., Bouma, T.J., Morris, E.P., Duarte, C.M., 2010. Effects of seagrasses and algae of the
 1057 *Caulerpa* family on hydrodynamics and particle-trapping rates. Mar. Biol. 157, 473-481.
 1058 Hernandez, A.L.M., van Tussenbroek, B.I., 2014. Patch dynamics and species shifts in seagrass
 1059 communities under moderate and high grazing pressure by green sea turtles. Mar. Ecol.-
 1060 Prog. Ser. 517, 143-157.
 1061 Hernandez, M.C., Sacco, J., Walsh, G.C., 2011. Biology and host preference of the planthopper *Taosa*
 1062 *longula* (Hemiptera: Dictyopharidae), a candidate for biocontrol of water hyacinth. Biocontrol
 1063 Sci. Techn. 21, 1079-1090.
 1064 Hidding, B., Bakker, E.S., Hootsmans, M.J.M., Hilt, S., 2016. Synergy between shading and herbivory
 1065 triggers macrophyte loss and regime shifts in aquatic systems. Oikos, DOI:
 1066 10.1111/oik.03104.
 1067 Hidding, B., Bakker, E.S., Keuper, F., de Boer, T., de Vries, P.P., Nolet, B.A., 2010. Differences in
 1068 tolerance of pondweeds and charophytes to vertebrate herbivores in a shallow Baltic
 1069 estuary. Aquat. Bot. 93: 123-128.
 1070 Hidding, B., Meirmans, P.G., Klaassen, M., de Boer, T., Ouborg, N.J., Wagemaker, C.A.M., Nolet, B.A.,
 1071 2014. The effect of herbivores on genotypic diversity in a clonal aquatic plant. Oikos 123,
 1072 1112-1120.
 1073 Hidding, B., Nolet, B.A., de Boer, T., de Vries, P.P., Klaassen, M., 2010. Above- and below-ground
 1074 vertebrate herbivory may each favour a different subordinate species in an aquatic plant
 1075 community. Oecologia 162, 199-208.
 1076 Hik, D.S., Sadul, H.A., Jefferies, R.L., 1991. Effects of the timing of multiple grazings by geese on net
 1077 aboveground primary production of swards of *Puccinellia phryganodes*. J. Ecol. 79, 715-730.
 1078 Hilt, S., 2006. Recovery of *Potamogeton pectinatus* L. stands in a shallow eutrophic lake under
 1079 extreme grazing pressure. Hydrobiologia 570, 95-99.
 1080 Hilt, S., 2015. Regime shifts between macrophytes and phytoplankton - concepts beyond shallow
 1081 lakes, unravelling stabilizing mechanisms and practical consequences. Limnetica 34, 467-480.

1082 Hilt, S., Henschke, I., Rucker, J., Nixdorf, B., 2010. Can submerged macrophytes influence turbidity
1083 and trophic state in deep lakes? Suggestions from a case study. J. Environ. Qual. 39, 725-733.

1084 Hilt, S., Kohler, J., Adrian, R., Monaghan, M.T., Sayer, C.D., 2013. Clear, crashing, turbid and back -
1085 long-term changes in macrophyte assemblages in a shallow lake. Freshw. Biol. 58, 2027-2036.

1086 Hilt, S., Kohler, J., Kozerski, H.P., van Nes, E.H., Scheffer, M., 2011. Abrupt regime shifts in space and
1087 time along rivers and connected lake systems. Oikos 120, 766-775.

1088 Hootsmans, M.J.M., 1999. Modelling *Potamogeton pectinatus*: for better or for worse. Hydrobiologia
1089 415, 7-11.

1090 Jackson, J.B.C., 1997. Reefs since Columbus. Coral Reefs 16, S23-S32.

1091 Jackson, M.C., Grey, J., 2013. Accelerating rates of freshwater invasions in the catchment of the River
1092 Thames. Biol. Invas. 15, 945-951.

1093 Jacobs, R.P.W.M., Den Hartog, C., Braster, B.F., Carriere, F.C. 1981. Grazing of the seagrass *Zostera*
1094 *noltii* by birds at Terschelling (Dutch Wadden Sea). Aquat. Bot. 10, 241-259.

1095 Jäger, P., Pall, K., Dumfahrt, E., 2004. A method of mapping macrophytes in large lakes with regard to
1096 the requirements of the Water Framework Directive. Limnologica 34, 140-146.

1097 Janzen, D.H., 1984. Dispersal of small seeds by big herbivores: foliage is the fruit. Am. Nat. 123, 338-
1098 353.

1099 Jefferies, R.L., Rockwell, R.F., Abraham, K.F., 2003. The embarrassment of riches: agricultural food
1100 subsidies, high goose numbers, and loss of Arctic wetlands - a continuing saga. Environ. Rev.
1101 11, 193-232.

1102 Jeppesen, E., Søndergaard, M., Jensen, J.P., Havens, K.E., Anneville, O., Carvalho, L., Coveney, M.F.,
1103 Deneke, R., Dokulil, M.T., Foy, B., Gerdeaux, D., Hampton, S.E., Hilt, S., Kangur, K., Kohler, J.,
1104 Lammens, E., Lauridsen, T.L., Manca, M., Miracle, M.R., Moss, B., Noges, P., Persson, G.,
1105 Phillips, G., Portielje, R., Schelske, C.L., Straile, D., Tatrai, I., Willen, E., Winder, M., 2005. Lake
1106 responses to reduced nutrient loading - an analysis of contemporary long-term data from 35
1107 case studies. Freshw. Biol. 50, 1747-1771.

1108 Jeppesen, E., Søndergaard, M., Søndergaard, M., Christoffersen, K., 1998, The structuring role of
 1109 submerged macrophytes in lakes. Springer.
 1110 Jessen, R.L., 1970. Mallard population trends and hunting losses in Minnesota. J. Wildlife Manage. 34,
 1111 93-105.
 1112 Jonzén, N., Nolet, B.A., Santamaría, L., Svensson, M.G.E., 2002. Seasonal herbivory and mortality
 1113 compensation in a swan- pondweed system. Ecol. Model. 147, 209-219.
 1114 Jupp, B.J., Spence, D.H.N., 1977. Limitations of macrophytes in a eutrophic lake, Loch Leven: II. Wave
 1115 action, sediments and waterfowl grazing. J. Ecol. 65, 431-446.
 1116 Kelkar, N., Arthur, R., Marba, N., Alcoverro, T., 2013a. Green turtle herbivory dominates the fate of
 1117 seagrass primary production in the Lakshadweep islands (Indian Ocean). Mar. Ecol.-Prog. Ser.
 1118 485, 235-243.
 1119 Kelkar, N., Arthur, R., Marba, N., Alcoverro, T., 2013b. Greener pastures? High-density feeding
 1120 aggregations of green turtles precipitate species shifts in seagrass meadows. J. Ecol. 101,
 1121 1158-1168.
 1122 Kerbes, R.H., Kotanen, P.M., Jefferies, R.L., 1990. Destruction of wetland habitats by Lesser Snow
 1123 Geese - A keystone species on the west-coast of Hudson-Bay. J. Appl. Ecol. 27, 242-258.
 1124 King, R.A., Gornall, R.J., Preston, C.D., Croft, J.M., 2002. Population differentiation of *Potamogeton*
 1125 *pectinatus* in the Baltic Sea with reference to waterfowl dispersal. Mol. Ecol. 11, 1947-1956.
 1126 Kirsch, K.D., Valentine, J.F., Heck, K.L., 2002. Parrotfish grazing on turtlegrass *Thalassia testudinum*:
 1127 evidence for the importance of seagrass consumption in food web dynamics of the Florida
 1128 Keys National Marine Sanctuary. Mar. Ecol.-Prog. Ser. 227, 71-85.
 1129 Kitchener, A.C., Conroy, J.W.H., 1997. The history of the Eurasian beaver *Castor fiber* in Scotland.
 1130 Mamm. Rev. 27, 95-108.
 1131 Klaassen, R.H.G., Nolet, B.A., Bankert, D., 2006. Movement of foraging Tundra Swans explained by
 1132 spatial pattern in cryptic food densities. Ecology 87, 2244-2254.

- 1133 Kleyheeg, E., 2015, Seed dispersal by a generalist duck: ingestion, digestion and transportation by
1134 mallards (*Anas platyrhynchos*). University of Utrecht.
- 1135 Körner, S., Dugdale, T., 2003. Is roach herbivory preventing re-colonization of submerged
1136 macrophytes in a shallow lake? *Hydrobiologia* 506, 497-501.
- 1137 Körner, S., Schreiber, E.S.G., Walz, N., 2002. Herbivory on submerged macrophytes - who's to blame?
1138 *Verh. Internat. Ver. Limnol.* 28, 967-970.
- 1139 Kuijper, D.P.J., de Kleine, C., Churski, M., van Hooft, P., Bubnicki, J., Jedrzejewska, B., 2013. Landscape
1140 of fear in Europe: wolves affect spatial patterns of ungulate browsing in Bialowieza Primeval
1141 Forest, Poland. *Ecography* 36, 1263-1275.
- 1142 Lal, A., Arthur, R., Marba, N., Lill, A.W.T., Alcoverro, T., 2010. Implications of conserving an ecosystem
1143 modifier: Increasing green turtle (*Chelonia mydas*) densities substantially alters seagrass
1144 meadows. *Biol. Conserv.* 143, 2730-2738.
- 1145 Laubek, B., 1995. Habitat use by Whooper Swans *Cygnus cygnus* and Bewick's Swans *Cygnus*
1146 *columbianus bewickii* wintering in Denmark: increasing agricultural conflicts. *Wildfowl* 46, 8-
1147 15.
- 1148 Lauridsen, T.L., Jeppesen, E., Andersen, F.O., 1993. Colonization of submerged macrophytes in
1149 shallow fish manipulated Lake Væng: impact of sediment composition and waterfowl grazing.
1150 *Aquat. Bot.* 46, 1-15.
- 1151 Letson, M.A., Makarewicz, J.C., 1994. An experimental test of the crayfish (*Orconectes immunis*) as a
1152 control mechanism for submersed aquatic macrophytes. *Lake Reserv. Manage.* 10, 127-132.
- 1153 Lodge, D.M., 1991. Herbivory on fresh-water macrophytes. *Aquat. Bot.* 41, 195-224.
- 1154 Lodge, D.M., Cronin, G., Van Donk, E., Froelich, A.J., 1998, Impact of herbivory on plant standing crop:
1155 comparisons among biomes, between vascular and non-vascular plants, and among
1156 freshwater herbivore taxa. In: Jeppesen, E., Søndergaard, M., Christoffersen, K. (Eds.), *The*
1157 *structuring role of submerged macrophytes in lakes*. Springer-Verlag, New York, pp. 149-174.

1158 Louette, G., De Meester, L., 2005. High dispersal capacity of cladoceran zooplankton in newly
 1159 founded communities. *Ecology* 86, 353-359.
 1160 Macreadie, P.I., Trevathan-Tackett, S.M., Skilbeck, C.G., Sanderman, J., Curlevski, N., Jacobsen, G.,
 1161 Seymour, J.R., 2015. Losses and recovery of organic carbon from a seagrass ecosystem
 1162 following disturbance. *P. R. Soc. B-Biol. Sci.* 282, 20151537.
 1163 Mader, E., van Vierssen, W., Schwenk, K., 1998. Clonal diversity in the submerged macrophyte
 1164 *Potamogeton pectinatus* L. inferred from nuclear and cytoplasmic variation. *Aquat. Bot.* 62,
 1165 147-160.
 1166 Madin, E.M.P., Madin, J.S., Booth, D.J., 2011. Landscape of fear visible from space. *Scientific Reports*
 1167 1, 14.
 1168 Mahdy, A., Hilt, S., Filiz, N., Beklioglu, M., Hejzlar, J., Ozkundakci, D., Papastergiadou, E.,
 1169 Scharfenberger, U., Sorf, M., Stefanidis, K., Tuvikene, L., Zingel, P., S ndergaard, M.,
 1170 Jeppesen, E., Adrian, R., 2015. Effects of water temperature on summer periphyton biomass
 1171 in shallow lakes: a pan-European mesocosm experiment. *Aquat. Sci.* 77, 499-510.
 1172 Mao, Z.G., Gu, X.H., Zeng, Q.F., Gu, X.K., Li, X.G., Wang, Y.P., 2014. Production sources and food web
 1173 of a macrophyte-dominated region in Lake Taihu, based on gut contents and stable isotope
 1174 analyses. *J. Great Lakes Res.* 40, 656-665.
 1175 Marklund, O., Sandsten, H., Hansson, L.A., Blindow, I., 2002. Effects of waterfowl and fish on
 1176 submerged vegetation and macroinvertebrates. *Freshw. Biol.* 47, 2049-2059.
 1177 McCauley, D.J., Pinsky, M.L., Palumbi, S.R., Estes, J.A., Joyce, F.H., Warner, R.R., 2015. Marine
 1178 defaunation: Animal loss in the global ocean. *Science* 347, 247-+.
 1179 McNaughton, 1985. Ecology of a Grazing Ecosystem: The Serengeti. *Ecol. Monogr.* 55, 260-294.
 1180 Mendonca, R., Kosten, S., Lacerot, G., Mazzeo, N., Roland, F., Ometto, J.P., Paz, E.A., Bove, C.P.,
 1181 Bueno, N.C., Gomes, J.H.C., Scheffer, M., 2013. Bimodality in stable isotope composition
 1182 facilitates the tracing of carbon transfer from macrophytes to higher trophic levels.
 1183 *Hydrobiologia* 710, 205-218.

1184 Miler, O., Straile, D., 2010. How to cope with a superior enemy? Plant defence strategies in response
1185 to annual herbivore outbreaks. *J. Ecol.* 98, 900-907.

1186 Mitchell, S.F., Wass, R.T., 1996a. Grazing by black swans (*Cygnus atratus* Latham), physical factors,
1187 and the growth and loss of aquatic vegetation in a shallow lake. *Aquat. Bot.* 55, 205-215.

1188 Mitchell, S.F., Wass, R.T., 1996b. Quantifying herbivory: Grazing consumption and interaction
1189 strength. *Oikos* 76, 573-576.

1190 Moles, A.T., Bonser, S.P., Poore, A.G.B., Wallis, I.R., Foley, W.J., 2011. Assessing the evidence for
1191 latitudinal gradients in plant defence and herbivory. *Funct. Ecol.* 25, 380-388.

1192 Moran, K.L., Bjorndal, K.A., 2005. Simulated green turtle grazing affects structure and productivity of
1193 seagrass pastures. *Mar. Ecol. Prog. Ser.* 305, 235-247.

1194 Morrison, W.E., Hay, M.E., 2012. Are lower-latitude plants better defended? Palatability of
1195 freshwater macrophytes. *Ecology* 93, 65-74.

1196 Moss, B., 2015. Mammals, freshwater reference states, and the mitigation of climate change.
1197 *Freshw. Biol.* 60, 1964-1976.

1198 Mueller, M.H., van der Valk, A.G., 2002. The potential role of ducks in wetland seed dispersal.
1199 *Wetlands* 22, 170-178.

1200 Munoz, J., Amat, F., Green, A.J., Figuerola, J., Gomez, A., 2013. Bird migratory flyways influence the
1201 phylogeography of the invasive brine shrimp *Artemia franciscana* in its native American
1202 range. *Peerj* 1, UNSP e200.

1203 Newman, R.M., 1991. Herbivory and detritivory on freshwater macrophytes by invertebrates - a
1204 review. *J. N. Am. Benth. Soc.* 10, 89-114.

1205 Newman, R.M., 2004. Biological control of Eurasian watermilfoil by aquatic insects: basic insights
1206 from an applied problem. *Arch. Hydrobiol.* 159, 145-184.

1207 Nichols, T.C., 2014. Ten years of resident Canada Goose damage management in a New Jersey tidal
1208 freshwater wetland. *Wildlife Soc. Bull.* 38, 221-228.

1209 Nolet, B.A., 2004. Overcompensation and grazing optimisation in a swan-pondweed system? Freshw.
 1210 Biol. 49, 1391-1399.

1211 Nolet, B.A., Gyimesi, A., Klaassen, R.H.G., 2006. Prediction of bird-day carrying capacity on a staging
 1212 site: a test of depletion models. J. Anim. Ecol. 75, 1285-1292.

1213 Nolet, B.A., Gyimesi, A., van Krimpen, R.R.D., de Boer, W.F., Stillman, R.A., 2016. Predicting effects of
 1214 water regime changes on waterbirds: insights from staging swans. PLoS One 11, e0147340.

1215 Nolet, B.A., Langevoord, O., Bevan, R.M., Engelaar, K.R., Klaassen, M., Mulder, R.J.W., Van Dijk, S.,
 1216 2001. Spatial variation in tuber depletion by swans explained by differences in net intake
 1217 rates. Ecology 82, 1655-1667.

1218 Nolet, B.A., Rosell, F., 1998. Comeback of the beaver *Castor fiber*: An overview of old and new
 1219 conservation problems. Biol. Conserv. 83, 165-173.

1220 Nolte, S., Esselink, P., Bakker, J.P., Smit, C., 2015. Effects of livestock species and stocking density on
 1221 accretion rates in grazed salt marshes. Estuar. Coast. Shelf S. 152, 109-115.

1222 Nummi, P., Holopainen, S., 2014. Whole-community facilitation by beaver: ecosystem engineer
 1223 increases waterbird diversity. Aquat. Conserv. 24, 623-633.

1224 Olsen, Y.S., Valiela, I., 2010. Effect of sediment nutrient enrichment and grazing on Turtle Grass
 1225 *Thalassia testudinum* in Jobos Bay, Puerto Rico. Estuaries Coasts 33, 769-783.

1226 Owen-Smith, R.N., 1988, Megaherbivores: the influence of very large body size on ecology.
 1227 Cambridge University Press, Cambridge.

1228 Pages, J.F., Farina, S., Gera, A., Arthur, R., Romero, J., Alcoverro, T., 2012. Indirect interactions in
 1229 seagrasses: fish herbivores increase predation risk to sea urchins by modifying plant traits.
 1230 Funct. Ecol. 26, 1015-1023.

1231 Pages, J.F., Gera, A., Romero, J., Alcoverro, T., 2014. Matrix composition and patch edges influence
 1232 plant-herbivore interactions in marine landscapes. Funct. Ecol. 28, 1440-1448.

1233 Paice, R.L., Chambers, J.M., Robson, B.J., 2016. Outcomes of submerged macrophyte restoration in a
 1234 shallow impounded, eutrophic river. Hydrobiologia DOI 10.1007/s10750-015-2441-8.

1235 Panov, V.E., Caceres, C., 2007. Role of diapause in dispersal of aquatic invertebrates. Diapause in
1236 Aquatic Invertebrates Theory and Human Use. Springer, Netherlands.

1237 Perry, D., Perry, G., 2008. Improving interactions between animal rights groups and conservation
1238 biologists. *Conserv. Biol.* 22, 27-35.

1239 Petruzzella, A., Guariento, R.D., Gripp, A.d.R., Marinho, C.C., Figueiredo-Barros, M.P., Esteves, F.d.A.,
1240 2015. Herbivore damage increases methane emission from emergent aquatic macrophytes.
1241 *Aquat. Bot.* 127, 6-11.

1242 Polunin, N.V.C., 1984. The decomposition of emergent macrophytes in freshwater. *Advances in Ecol.*
1243 *Res.* 14, 115-166.

1244 Poore, A.G.B., Campbell, A.H., Coleman, R.A., Edgar, G.J., Jormalainen, V., Reynolds, P.L., Sotka, E.E.,
1245 Stachowicz, J.J., Taylor, R.B., Vanderklift, M.A., Duffy, J.E., 2012. Global patterns in the impact
1246 of marine herbivores on benthic primary producers. *Ecol. Lett.* 15, 912-922.

1247 Prado, P., Romero, J., Alcoverro, T., 2010. Nutrient status, plant availability and seasonal forcing
1248 mediate fish herbivory in temperate seagrass beds. *Mar. Ecol.-Prog. Ser.* 409, 229-239.

1249 Prado, P., Tomas, F., Alcoverro, T., Romero, J., 2007. Extensive direct measurements of *Posidonia*
1250 *oceanica* defoliation confirm the importance of herbivory in temperate seagrass meadows.
1251 *Mar. Ecol.-Prog. Ser.* 340, 63-71.

1252 Preen, A., 1995. Impacts of Dugong foraging on seagrass habitats - observational and experimental
1253 evidence for cultivation grazing. *Mar. Ecol.-Prog. Ser.* 124, 201-213.

1254 Prejs, A., 1984. Herbivory by temperate freshwater fishes and its consequences. *Environ. Biol. Fish.*
1255 10, 281-296.

1256 Ramo, C., Amat, J.A., Nilsson, L., Schricke, V., Rodriguez-Alonso, M., Gomez-Crespo, E., Jubete, F.,
1257 Navedo, J.G., Masero, J.A., Palacios, J., Boos, M., Green, A.J., 2015. Latitudinal-related
1258 variation in wintering population trends of greylag geese (*Anser anser*) along the Atlantic
1259 flyway: a response to climate change? *PLoS One* 10, e0140181.

1260 Redpath, S.M., Gutierrez, R.J., Wood, K.A., Young, J.C., 2015. Conflicts in conservation: navigating
 1261 towards solutions. Cambridge University Press, Cambridge, UK.

1262 Reynolds, C., Miranda, N.A.F., Cumming, G.S., 2015. The role of waterbirds in the dispersal of aquatic
 1263 alien and invasive species. *Divers. Distrib.* 21, 744-754.

1264 Robledo-Arnuncio, J.J., Klein, E.K., Muller-Landau, H.C., Santamaría, L., 2014. Space, time and
 1265 complexity in plant dispersal ecology. *Movement Ecol.* 2, 16.

1266 Röckstrom, J., Steffen, W., Noone, K., Persson, A., Chapin, F.S., Lambin, E.F., Lenton, T.M., Scheffer,
 1267 M., Folke, C., Schellnhuber, H.J., Nykvist, B., de Wit, C.A., Hughes, T., van der Leeuw, S.,
 1268 Rodhe, H., Sorlin, S., Snyder, P.K., Costanza, R., Svedin, U., Falkenmark, M., Karlberg, L.,
 1269 Corell, R.W., Fabry, V.J., Hansen, J., Walker, B., Liverman, D., Richardson, K., Crutzen, P.,
 1270 Foley, J.A., 2009. A safe operating space for humanity. *Nature* 461, 472-475.

1271 Rodriguez, C.F., Becares, E., Fernandez-Alaez, M., 2003. Shift from clear to turbid phase in Lake
 1272 Chozas (NW Spain) due to the introduction of American red swamp crayfish (*Procambarus*
 1273 *clarkii*). *Hydrobiologia* 506, 421-426.

1274 Ross, E., 1971. Biological control of pond weeds with Chinese Geese. *Hawaii Farm Sci.* 20, 11-12.

1275 Rushing, W.N., 1973. Water hyacinth research in Puerto Rico. *Hyacinth Contr. J.* 13, 48-54.

1276 Sachse, R., Petzoldt, T., Blumstock, M., Moreira, S., Patzig, M., Rueker, J., Janse, J.H., Mooij, W.M.,
 1277 Hilt, S., 2014. Extending one-dimensional models for deep lakes to simulate the impact of
 1278 submerged macrophytes on water quality. *Environ. Modell. Softw.* 61, 410-423.

1279 Sand-Jensen, K., Jacobsen, D., 2002. Herbivory and growth in terrestrial and aquatic populations of
 1280 amphibious stream plants. *Freshw. Biol.* 47, 1475-1487.

1281 Sand-Jensen, K., Jacobsen, D., Duarte, C.M., 1994. Herbivory and resulting plant damage. *Oikos* 69,
 1282 545-549.

1283 Sandom, C.J., Ejrnaes, R., Hansen, M.D.D., Svenning, J.C., 2014. High herbivore density associated
 1284 with vegetation diversity in interglacial ecosystems. *P. Natl. Acad. Sci. USA* 111, 4162-4167.

1285 Santamaria, L., Charalambidou, I., Figuerola, J., Green, A.J., 2002. Effect of passage through duck gut
1286 on germination of fennel pondweed seeds. *Arch. Hydrobiol.* 156, 11-22.

1287 Sarneel, J.M., Huig, N., Veen, G.F., Rip, W., Bakker, E.S., 2014. Herbivores enforce sharp boundaries
1288 between terrestrial and aquatic ecosystems. *Ecosystems* 17, 1426-1438.

1289 Scharnweber, K., Syvaranta, J., Hilt, S., Brauns, M., Vanni, M.J., Brothers, S., Kohler, J., Knezevic-Jaric,
1290 J., Mehner, T., 2014. Whole-lake experiments reveal the fate of terrestrial particulate organic
1291 carbon in benthic food webs of shallow lakes. *Ecology* 95, 1496-1505.

1292 Scheffer, M., Hosper, S.H., Meijer, M.-L., Moss, B., Jeppesen, E., 1993. Alternative equilibria in
1293 shallow lakes. *Trends Ecol. Evol.* 8, 275-279.

1294 Schemske, D.W., Mittelbach, G.G., Cornell, H.V., Sobel, J.M., Roy, K., 2009. Is there a latitudinal
1295 gradient in the importance of biotic interactions? *Annu. Rev. Ecol. Evol. S.* 40, 245-269.

1296 Schwartz, D.P., Maughan, O.E., Gebhart, G.E., 1986. Effectiveness of Blue Tilapia as a weed-control
1297 agent in ponds. *Progressive Fish-Culturist* 48, 259-263.

1298 Shelford, V.E., 1918, Conditions of existence. In: Ward, H.B., Whipple, G.C. (Eds.), *Freshwater Biology*.
1299 John Wiley, New York, pp. 21-60.

1300 Sherfy, M.H., Kirkpatrick, R.L., 2003. Invertebrate response to snow goose herbivory on moist-soil
1301 vegetation. *Wetlands* 23, 236-249.

1302 Silva, T.S.F., Costa, M.P.F., Melack, J.M., Novo, E.M.L.M., 2008. Remote sensing of aquatic vegetation:
1303 theory and applications. *Environ. Monit. Assess.* 140, 131-145.

1304 Skilleter, G.A., Wegscheidl, C., Lanyon, J.M., 2007. Effects of grazing by a marine mega-herbivore on
1305 benthic assemblages in a subtropical seagrass bed. *Mar. Ecol. Prog. Ser.* 351, 287-300.

1306 Small, E., 2012. The new Noah's Ark: beautiful and useful species only. Part 2. The chosen species.
1307 *Biodiversity* 13, 37-53.

1308 Smolders, A.J.P., Vergeer, L.H.T., van der Velde, G., Roelofs, J.G.M., 2000. Phenolic contents of
1309 submerged, emergent and floating leaves of aquatic and semi-aquatic macrophyte species:
1310 why do they differ? *Oikos* 91, 307-310.

1311 Solomon, C.T., Carpenter, S.R., Clayton, M.K., Cole, J.J., Coloso, J.J., Pace, M.L., Vander Zanden, M.J.,
 1312 Weidel, B.C., 2011. Terrestrial, benthic, and pelagic resource use in lakes: results from a
 1313 three-isotope Bayesian mixing model. *Ecology* 92, 1115-1125.
 1314 Søndergaard, M., Bruun, L., Lauridsen, T., Jeppesen, E., Madsen, T.V., 1996. The impact of grazing
 1315 waterfowl on submerged macrophytes: In situ experiments in a shallow eutrophic lake.
 1316 *Aquat. Bot.* 53, 73-84.
 1317 Søndergaard, M., Jeppesen, E., Lauridsen, T.L., Skov, C., Van Nes, E.H., Roijackers, R., Lammens, E.,
 1318 Portielje, R., 2007. Lake restoration: successes, failures and long-term effects. *J. Appl. Ecol.*
 1319 44, 1095-1105.
 1320 Soons, M.B., van der Vlugt, C., van Lith, B., Heil, G.W., Klaassen, M., 2008. Small seed size increases
 1321 the potential for dispersal of wetland plants by ducks. *J. Ecol.* 96, 619-627.
 1322 Soti, P.G., Volin, J.C., 2010. Does water hyacinth (*Eichhornia crassipes*) compensate for simulated
 1323 defoliation? Implications for effective biocontrol. *Biol. Control* 54, 35-40.
 1324 Stillman, R.A., Wood, K.A., Gilkerson, W., Elkinton, E., Black, J.M., Ward, D.H., Petrie, M., 2015.
 1325 Predicting effects of environmental change on a migratory herbivore. *Ecosphere* 6, art.114.
 1326 Stott, B., Robson, T.O., 1970. Efficiency of grass carp (*Ctenopharyngodon idella* Val) in controlling
 1327 submerged water weeds. *Nature* 226, 870-&.
 1328 Subalusky, A.L., Dutton, C.L., Rosi-Marshall, E.J., Post, D.M., 2015. The hippopotamus conveyor belt:
 1329 vectors of carbon and nutrients from terrestrial grasslands to aquatic systems in sub-Saharan
 1330 Africa. *Freshw. Biol.* 60, 512-525.
 1331 Sumoski, S.E., Orth, R.J., 2012. Biotic dispersal in eelgrass *Zostera marina*. *Mar. Ecol.-Prog. Ser.* 471,
 1332 1-10.
 1333 Tatu, K.S., Anderson, J.T., Hindman, L.J., Seidel, G., 2007. Mute swans' impact on submerged aquatic
 1334 vegetation in Chesapeake Bay. *J. Wildlife Manage.* 71, 1431-1439.
 1335 Thayer, G.W., Engel, D.W., Bjorndal, K.A., 1982. Evidence for short-circuiting of the detritus cycle of
 1336 seagrass beds by the green turtle, *Chelonia mydas*. *J. Exp. Mar. Biol. Ecol.* 62, 173-183.

1337 Tomas, F., Turon, X., Romero, J., 2005. Seasonal and small-scale spatial variability of herbivory
1338 pressure on the temperate seagrass *Posidonia oceanica*. Mar. Ecol.-Prog. Ser. 301, 95-107.

1339 Turcotte, M.M., Davies, T.J., Thomsen, C.J.M., Johnson, M.T.J., 2014. Macroecological and
1340 macroevolutionary patterns of leaf herbivory across vascular plants. P. R. Soc. B-Biol. Sci.-
1341 Biological Sciences 281, 20140555.

1342 Turvey, S.T., Risley, C.L., 2006. Modelling the extinction of Steller's sea cow. Biol. Lett. 2, 94-97.

1343 Vadeboncoeur, Y., Vander Zanden, M.J., Lodge, D.M., 2002. Putting the lake back together:
1344 Reintegrating benthic pathways into lake food web models. Bioscience 52, 44-54.

1345 Valentine, J.F., Duffy, J.E., 2006, The central role of grazing in seagrass ecology. In: Larkum, A.W.D.,
1346 Orth, R.J., Duarte, C.M. (Eds.), Seagrasses: Biology, Ecology and Conservation. Springer, pp.
1347 463-501.

1348 Valentine, J.F., Heck, K.L., 1991. The role of sea-urchin grazing in regulating subtropical seagrass
1349 meadows - evidence from field manipulations in the northern Gulf of Mexico. J. Exp. Mar.
1350 Biol. Ecol. 154, 215-230.

1351 Valentine, J.F., Heck, K.L., 1999. Seagrass herbivory: evidence for the continued grazing of marine
1352 grasses. Mar. Ecol.-Prog. Ser. 176, 291-302.

1353 Valentine, J.F., Heck, K.L., Busby, J., Webb, D., 1997. Experimental evidence that herbivory increases
1354 shoot density and productivity in a subtropical turtlegrass (*Thalassia testudinum*) meadow.
1355 Oecologia 112, 193-200.

1356 Van der Pijl, L., 1982, Principles of dispersal. Springer-Verlag, Berlin.

1357 Van der Wal, J.E.M., Dorenbosch, M., Immers, A.K., Forteza, C.V., Geurts, J.J.M., Peeters, E., Koese, B.,
1358 Bakker, E.S., 2013. Invasive crayfish threaten the development of submerged macrophytes in
1359 lake restoration. PLoS One 8, e78579.

1360 Van der Wal, R., Sjogersten, S., Woodin, S.J., Cooper, E.J., Jonsdottir, I.S., Kuijper, D., Fox, T.A.D.,
1361 Huiskes, A.D., 2007. Spring feeding by pink-footed geese reduces carbon stocks and sink
1362 strength in tundra ecosystems. Global Change Biology 13, 539-545.

1363 Van Donk, E., Otte, A., 1996. Effects of grazing by fish and waterfowl on the biomass and species
 1364 composition of submerged macrophytes. *Hydrobiologia* 340, 285-290.

1365 Van Eerden, M.R., Drent, R.H., Stahl, J., Bakker, J.P., 2005. Connecting seas: western Palaearctic
 1366 continental flyway for water birds in the perspective of changing land use and climate. *Glob.*
 1367 *Change Biol.* 11, 894-908.

1368 Van Leeuwen, C.H.A., Huig, N., Van der Velde, G., Van Alen, T.A., Wagemaker, C.A.M., Sherman,
 1369 C.D.H., Klaassen, M., Figuerola, J., 2013. How did this snail get here? Several dispersal vectors
 1370 inferred for an aquatic invasive species. *Freshw. Biol.* 58, 88-99.

1371 Van Leeuwen, C.H.A., Van der Velde, G., van Lith, B., Klaassen, M., 2012. Experimental quantification
 1372 of long distance dispersal potential of aquatic snails in the gut of migratory birds. *PLoS One* 7,
 1373 e32292.

1374 Van Nes, E.H., Scheffer, M., van den Berg, M.S., Coops, H., 2003. Charisma: a spatial explicit
 1375 simulation model of submerged macrophytes. *Ecol. Modell.* 159, 103-116.

1376 Van Wijk, R.J., 1988. Ecological studies on *Potamogeton pectinatus* L.I. General characteristics,
 1377 biomass production and life cycles under field conditions. *Aquat. Bot.* 31, 211-258.

1378 Vander Zanden, H.B., Bjorndal, K.A., Bolten, A.B., 2013. Temporal consistency and individual
 1379 specialization in resource use by green turtles in successive life stages. *Oecologia* 173, 767-
 1380 777.

1381 Veen, G.F., Sarneel, J.M., Ravensbergen, L., Huig, N., van Paassen, J., Rip, W., Bakker, E.S., 2013.
 1382 Aquatic grazers reduce the establishment and growth of riparian plants along an
 1383 environmental gradient. *Freshw. Biol.* 58, 1794-1803.

1384 Verges, A., Becerro, M.A., Alcoverro, T., Romero, J., 2007. Variation in multiple traits of vegetative
 1385 and reproductive seagrass tissues influences plant-herbivore interactions. *Oecologia* 151,
 1386 675-686.

1387 Verges, A., Perez, M., Alcoverro, T., Romero, J., 2008. Compensation and resistance to herbivory in
 1388 seagrasses: induced responses to simulated consumption by fish. *Oecologia* 155, 751-760.

1389 Verges, A., Steinberg, P.D., Hay, M.E., Poore, A.G.B., Campbell, A.H., Ballesteros, E., Heck, K.L., Booth,
 1390 D.J., Coleman, M.A., Feary, D.A., Figueira, W., Langlois, T., Marzinelli, E.M., Mizerek, T.,
 1391 Mumby, P.J., Nakamura, Y., Roughan, M., van Sebille, E., Sen Gupta, A., Smale, D.A., Tomas,
 1392 F., Wernberg, T., Wilson, S.K., 2014a. The tropicalization of temperate marine ecosystems:
 1393 climate-mediated changes in herbivory and community phase shifts. P. R. Soc. B-Biol. Sci.
 1394 281, 20140846.

1395 Verges, A., Tomas, F., Cebrian, E., Ballesteros, E., Kizilkaya, Z., Dendrinos, P., Karamanlidis, A.A.,
 1396 Spiegel, D., Sala, E., 2014b. Tropical rabbitfish and the deforestation of a warming temperate
 1397 sea. J. Ecol. 102, 1518-1527.

1398 Verhoeven, J.T.A., 1980. The ecology of *Ruppia*-dominated communities in western-Europe. 3.
 1399 Aspects of production, consumption and decomposition. Aquat. Bot. 8, 209-253.

1400 Verpoorter, C., Kutser, T., Seekell, D.A., Tranvik, L.J., 2014. A global inventory of lakes based on high-
 1401 resolution satellite imagery. Geophys. Res. Lett. 41, 6396-6402.

1402 Viana, D.S., Figuerola, J., Schwenk, K., Manca, M., Hobæk, A., Mjelde, M., Preston, C.D., Gornall, R.J.,
 1403 Croft, J.M., King, R.A., Green, A.J., Santamaría, L., 2016. Assembly mechanisms determining
 1404 high species turnover in aquatic communities over regional and continental scales. Ecography
 1405 39, 281-288.

1406 Viana, D.S., Santamaria, L., Michot, T.C., Figuerola, J., 2013a. Allometric scaling of long-distance seed
 1407 dispersal by migratory birds. Am. Nat. 181, 649-662.

1408 Viana, D.S., Santamaria, L., Michot, T.C., Figuerola, J., 2013b. Migratory strategies of waterbirds
 1409 shape the continental-scale dispersal of aquatic organisms. Ecography 36, 430-438.

1410 Viana, D.S., Santamaria, L., Schwenk, K., Manca, M., Hobaek, A., Mjelde, M., Preston, C.D., Gornall,
 1411 R.J., Croft, J.M., King, R.A., Green, A.J., Figuerola, J., 2014. Environment and biogeography
 1412 drive aquatic plant and cladoceran species richness across Europe. Freshw. Biol. 59, 2096-
 1413 2106.

1414 Vivian-Smith, G., Stiles, E.W., 1994. Dispersal of salt-marsh seeds on the feet and feathers of
 1415 waterfowl. *Wetlands* 14, 316-319.

1416 Vonk, J.A., Kneer, D., Stapel, J., Asmus, H., 2008a. Shrimp burrow in tropical seagrass meadows: An
 1417 important sink for litter. *Estuar. Coast. Shelf S.* 79, 79-85.

1418 Vonk, J.A., Pijnappels, M.H.J., Stapel, J., 2008b. In situ quantification of *Tripneustes gratilla* grazing
 1419 and its effects on three co-occurring tropical seagrass species. *Mar. Ecol. Prog. Ser.* 360, 107-
 1420 114.

1421 Weisz, E.J., Yan, N.D., 2010. Relative value of limnological, geographic, and human use variables as
 1422 predictors of the presence of *Bythotrephes longimanus* in Canadian Shield lakes. *Can. J. Fish.*
 1423 *Aquat. Sci.* 67, 462-472.

1424 Wheeler, G.S., Center, T.D., 2001. Impact of the biological control agent *Hydrellia pakistanae* (Diptera
 1425 : Ephydriidae) on the submersed aquatic weed *Hydrilla verticillata* (Hydrocharitaceae). *Biol.*
 1426 *Control* 21, 168-181.

1427 Wheeler, G.S., Van, T.K., Center, T.D., 1998. Herbivore adaptations to a low-nutrient food: Weed
 1428 biological control specialist *Spodoptera pectinicornis* (Lepidoptera : Noctuidae) fed the
 1429 floating aquatic plant *Pistia stratiotes*. *Environ. Entomol.* 27, 993-1000.

1430 Whitehead, P.J.P., 1978. Ancient records of the presence of the Caribbean manatee *Trichechus*
 1431 *manatus* in Brazil. *Acta Amazonica* 8, 497-506.

1432 Wilson, F., 1964. Biological control of weeds. *Annu. Rev. Entomol.* 9, 225-&.

1433 Wilson, H.R., Harms, R.H., Damron, B.L., 1977. Potential of geese in control and utilization of water
 1434 hyacinths. *Poultry Sci.* 56, 1360-1361.

1435 Winston, J.E., 2012. Dispersal in marine organisms without a pelagic larval phase. *Integr. Comp. Biol.*
 1436 52, 447-457.

1437 Wittmann, M.E., Jerde, C.L., Howeth, J.G., Maher, S.P., Deines, A.M., Jenkins, J.A., Whitledge, G.W.,
 1438 Burbank, S.R., Chadderton, W.L., Mahon, A.R., Tyson, J.T., Gantz, C.A., Keller, R.P., Drake,
 1439 J.M., Lodge, D.M., 2014. Grass carp in the Great Lakes region: establishment potential, expert

1440 perceptions, and re-evaluation of experimental evidence of ecological impact. *Can. J. Fish.*
 1441 *Aquat. Sci.* 71, 992-999.
 1442 Wood, K.A., O'Hare, M.T., McDonald, C., Searle, K.R., Daunt, F., Stillman, R.T., 2016. Herbivore
 1443 regulation of plant abundance in aquatic ecosystems. *Biological Reviews* DOI:
 1444 10.1111/brv.12272.
 1445 Wood, K.A., Stillman, R.A., Clarke, R.T., Daunt, F., O'Hare, M.T., 2012a. The impact of waterfowl
 1446 herbivory on plant standing crop: a meta-analysis. *Hydrobiologia* 686, 157-167.
 1447 Wood, K.A., Stillman, R.A., Clarke, R.T., Daunt, F., O'Hare, M.T., 2012b. Understanding plant
 1448 community responses to combinations of biotic and abiotic factors in different phases of the
 1449 plant growth cycle. *PLoS One* 7, e49824.
 1450 Wood, K.A., Stillman, R.A., Daunt, F., O'Hare, M.T., 2014a. Can sacrificial feeding areas protect
 1451 aquatic plants from herbivore grazing? Using behavioural ecology to inform wildlife
 1452 management. *PLoS One* 9, e104034.
 1453 Wood, K.A., Stillman, R.A., Daunt, F., O'Hare, M.T., 2014b. Chalk streams and grazing mute swans.
 1454 *British Wildlife* 25, 171-176.
 1455 Wood, K.A., Stillman, R.A., Daunt, F., O'Hare, M.T., 2015. The swan grazing conflict in chalk rivers. In:
 1456 Redpath, S.M., Gutierrez, R.J., Wood, K.A., Young, J.C. (Eds.), *Conflicts in conservation:*
 1457 *navigating towards solutions.* Cambridge University Press, Cambridge, UK, pp. 134-136.
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Captions to figures

Figure 1. Frequency diagram of percentage of herbivory (% herbivory) on vascular plants (a: mean percent damage; b, c: net primary production removal) and biomass C:N ratio (g/g) across (a,d) terrestrial, (b,e) freshwater and (c,f) marine ecosystems. Freshwater and marine data include both submerged and emergent plants. The median values in each panel are indicated with an arrow accompanied by “M”.

Data sources: terrestrial (a) herbivory: percentage of leaf area damaged (Turcotte et al. 2014), (d) biomass C:N ratio: in foliage (Elser et al. 2000); freshwater (b) herbivory: percentage of emergent and submerged vascular plant biomass removed by herbivores in herbivore exclosure/enclosure or addition/removal experiments (Wood et al. 2016), (e) biomass C:N ratio: in submerged and emergent vascular plants (Cloern et al. 2002; Bakker unpublished data); marine, (c) herbivory: percentage of emergent and submerged vascular plant biomass removed by herbivores in herbivore exclosure/enclosure or addition/removal experiments (Wood et al. 2016), percentage of leaf area damaged in seagrasses (Cebrian and Duarte 1998) (f) biomass C:N ratio: in seagrass leaves (Atkinson & Smith 1983, Duarte 1990, Fourqurean et al. 1993, Cebrian and Duarte 1998, Fourqurean et al. 2010, Olsen and Valiela 2010), in salt marsh plants (Cloern et al. 2002). The duration of the herbivory studies varied from instantaneous measurements of percentage leaf damage in terrestrial plants and seagrasses to exclosures studies in freshwater and marine ecosystems ranging from about a week to multiple study years (study durations reported in Wood et al. 2016 who found no effect of study duration on percentage herbivore plant biomass removal).

Figure 2. Frequency diagram of net primary production removal (% herbivory) and biomass C:N (g/g) for vascular freshwater and marine macrophytes of different growth forms (emergent and

1484 submerged). Emergent plants include floating plants and wetland or salt marsh plants. The median
1485 values in each panel are indicated with an arrow accompanied by “M”. Data sources as in Fig. 1.

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1487 Figure 3. Synthesizing scheme indicating the effects of herbivores on macrophyte beds and the
1488 functioning of shallow freshwater (a) and marine (b) aquatic ecosystems. Herbivores affect plant
1489 abundance and species composition by grazing and bioturbation. Their presence alters
1490 biogeochemical cycling and primary production, they transport nutrients and propagules across
1491 ecosystem boundaries, modify habitat for other organisms and affect the level of shoreline
1492 protection by macrophyte beds. Symbols in the figure are courtesy of the Integration and Application
1493 Network, Univ. of Maryland Center for Environmental Science (ian.umces.edu/symbols/).

1494 Table 1. Plant and herbivore traits promoting propagule dispersal by aquatic herbivores.

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Trait	Effect	References
<i>Herbivores</i>		
Ability to chew or grind food	The presence of a gizzard or grinding teeth reduces propagule survival. Among waterfowl, heavier gizzards reduce seed survival but higher grit content may enhance germination of undigested seeds.	Figuerola et al. (2002)
Furry or sticky appearance of animal body	Animals with a surface on which propagules can attach disperse more propagules	
Diet selection	Targeted feeding on seeds may result in more transport, but also in more seed predation – thus reducing the transport through untargeted feeding which, particularly when mixed with large bulks of food (plant parts, animal food, debris), may result in high propagule survival.	Figuerola et al. (2003)
Travelling distance	Larger travelling distances results in further potential dispersal, particularly for migratory species that cover long distances in single leaps	Viana et al. (2013b)
Habitat use	Animals with specialized use of aquatic habitats are more likely to deposit the propagules in suitable habitat. In particular, targeted arrival to aquatic habitats at stopovers may increased the deposition of	Figuerola and Green (2005)

propagules ingested at departure sites, especially after the first drinking and feeding bout.

Plants

Propagule dimensions	Small, round seeds survive digestive tract better than large, elongated seeds. Small size, in particular, may enhance ingestion mixed with vegetative plant material, increasing propagule ingestion (“foliage is the fruit”, sensu Janzen (1984)) and survival to gut passage.	Mueller and van der Valk (2002); Soons et al. (2008); Figuerola et al. (2010)
Hardness and permeability of seed coat	Thicker, harder and less permeable seed coats increase survival to disperser’s gut passage, but may reduce germination in the absence of uningested seeds.	Mueller and van der Valk (2002); Santamaria et al. (2002); Figuerola et al. (2010)
Adaptations for epizoochorous dispersal	Hooks, rough or sticky surface have been proposed to enhance dispersal potential	Van der Pijl (1982)
Resistance to dessication	Organism’s and/or propagule’s resistance to dessication may enhance epizoochorous dispersal of aquatic organisms	Panov and Caceres (2007); Havel et al. (2014)

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1499 Table 2. An overview of reported conservation conflicts that have arisen from the impacts of overgrazing by herbivores on macrophytes.

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Herbivore	Issue	Parties in conflict	Location(s)	Duration	Current status	References
Green turtle (<i>Chelonia mydas</i>)	Overgrazing of seagrasses can undermine conservation efforts in protected areas and reduce fish catches for local people	Turtle and seagrass conservationists, fishermen	Indo-Pacific oceans	1980s - present	Ongoing	Arthur et al. (2013); Christianen et al. (2014)
West Indian manatee (<i>Trichechus manatus</i>)	Overgrazing has hindered efforts to restore submerged macrophyte beds	Manatee and macrophyte conservationists	Freshwater and brackish ecosystems in south-east USA	1990s - present	Ongoing	Hauxwell et al. (2004)
Beaver (<i>Castor spp.</i>)	Impacts on aquatic habitats, via effects on vegetation and wider ecosystem (e.g. fish)	Conservationists, animal welfare groups, fishermen, and statutory wildlife management	Freshwater habitats across North America, Russia	Unknown - present	Ongoing	Nolet and Rosell (1998); Collen and Gibson (2001); Halley and Rosell (2002)

		agencies				
Coypu (<i>Myocastor coypus</i>) and muskrat (<i>Ondatra zibethicus</i>)	Overgrazing on emergent macrophytes degrades aquatic habitats	Conservationists, animal welfare groups, and statutory wildlife management agencies	Freshwater lakes and wetlands in Europe	1930s – present	Resolved by 1970s via extirpation of coypu in Britain; ongoing elsewhere in Europe	Gosling and Baker (1989); Barends (2002)
Mute swan (<i>Cygnus olor</i>)	Overgrazing of macrophytes degrades aquatic habitats	Conservationists, animal welfare groups, and statutory wildlife management agencies	Freshwater habitats in Europe and USA	1950s - present	Ongoing	Perry and Perry (2008); Wood et al. (2014b, 2015)
Geese (<i>Anser</i> spp., <i>Branta</i> spp., and <i>Chen</i> spp.)	Overgrazing of emergent macrophytes degrades wetland habitats	Conservationists, animal welfare groups, and statutory wildlife management agencies	Canadian Arctic and sub-Arctic wetlands, freshwater lakes	1970s - present	Ongoing	Kerbes et al. (1990); Nichols (2014)

agencies

and wetlands in

Europe and

North America

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1504 Table 3. A summary of key herbivore taxa used as biocontrol agents in the management of macrophytes.

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Herbivore biocontrol agent	Target macrophyte(s)	Herbivore generalist or specific?	References
West Indian manatee (<i>Trichechus manatus</i>)	Wide range of macrophyte species, including <i>Cabomba aquatica</i> , <i>Anacharis</i> spp., <i>Leersia</i> spp., <i>Utricularia</i> spp.	Generalist	Allsopp (1960)
Geese (<i>Anser</i> spp)	Wide range of macrophyte species	Generalist	Ross (1971); Wilson et al. (1977)
Grass carp (<i>Ctenopharyngodon idella</i>)	Wide range of macrophyte species, including invasive species such as <i>Hydrilla verticillata</i>	Generalist	Clayton (1996); Hanlon et al. (2000)
Cichlid fishes, e.g. blue tilapia (<i>Oreochromis aureus</i>)	Wide range of macrophyte species	Generalist	Schwartz et al. (1986)
Crayfish, e.g. papershell crayfish (<i>Orconectes immunis</i>)	Wide range of submerged macrophyte species	Generalist	Letson and Makarewicz (1994)
Weevils, e.g. milfoil weevil	Species-specific biocontrol agents identified for many macrophyte	Specialist	Creed and Sheldon

<i>(Euhrychiopsis lecontei)</i> , Hydrilla tuber weevil (<i>Bagous affinis</i>)	species, e.g. Hydrilla (<i>Hydrilla verticillata</i>), Eurasian water milfoil (<i>Myriophyllum spicatum</i>)		(1993); Newman (2004)
Apple snails (<i>Pomacea</i> spp.)	Wide range of macrophyte species	Generalist	Rushing (1973)
Dipteran larvae, e.g. Asian hydrilla leaf-mining fly (<i>Hydrellia pakistanae</i>)	Species-specific biocontrol agents identified for many macrophyte species, e.g. <i>Hydrilla verticillata</i>	Specialist	Wheeler and Center (2001); Bownes (2014)
Lepidopteran larvae, e.g. waterlettuce moth (<i>Spodoptera pectinicornis</i>)	Species-specific biocontrol agents identified for many macrophyte species	Both generalist and specialist species reported	Wheeler et al. (1998); Gross et al. (2001); Newman (2004)
Hemiptera, e.g. <i>Eccritotarsus catarinensis</i>	Species-specific biocontrol agents identified for many macrophyte species, e.g. Water hyacinth (<i>Eichhornia crassipes</i>)	Specialist	Coetzee et al. (2007); Hernandez et al. (2011)
Orthoptera, e.g. water hyacinth grasshopper (<i>Cornops aquaticum</i>)	Species-specific biocontrol agents identified for many macrophyte species, e.g. Water hyacinth (<i>Eichhornia crassipes</i>)	Specialist	Bownes et al. (2010)

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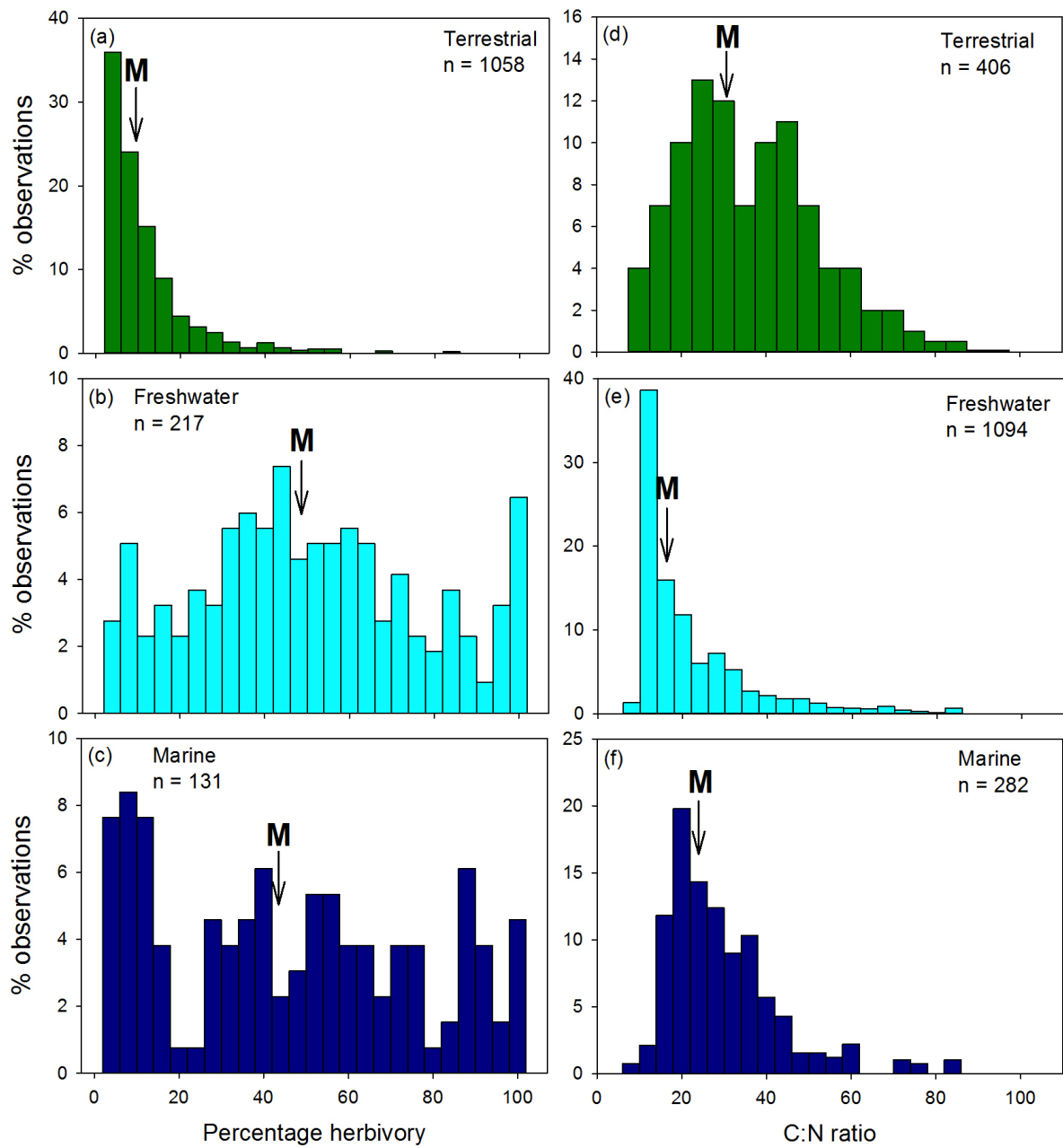
1508 Table 4. Different methodologies used to detect and quantify herbivory on macrophytes.

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Methodology	Explanation	References
Exclosures	Installation of cages (fully closed or open at the bottom) to protect macrophytes from different herbivores (fish, muskrats, waterfowl, crayfish, turtles)	Søndergaard et al. (1996); Körner et al. (2002); Hilt (2006); Christianen et al. (2012); Poore et al. (2012); Veen et al. (2013); Van der Wal et al. (2013); Sarneel et al. (2014)
Tethering	Shoot herbivory rates (cm shoot-1 day-1) are estimated for marked shoots by measuring leaf elongation over time.	Kirsch et al. (2002); Tomas et al. (2005); Prado et al. (2007); Pages et al. (2014)
Underwater videos	Video recording and quantification of fish activities including plucking of leaves	Körner & Dugdale (2003); Bennett & Bellwood (2011); Verges et al. (2014b)
Visual estimation of leaf damage		Francescini et al. (2010)

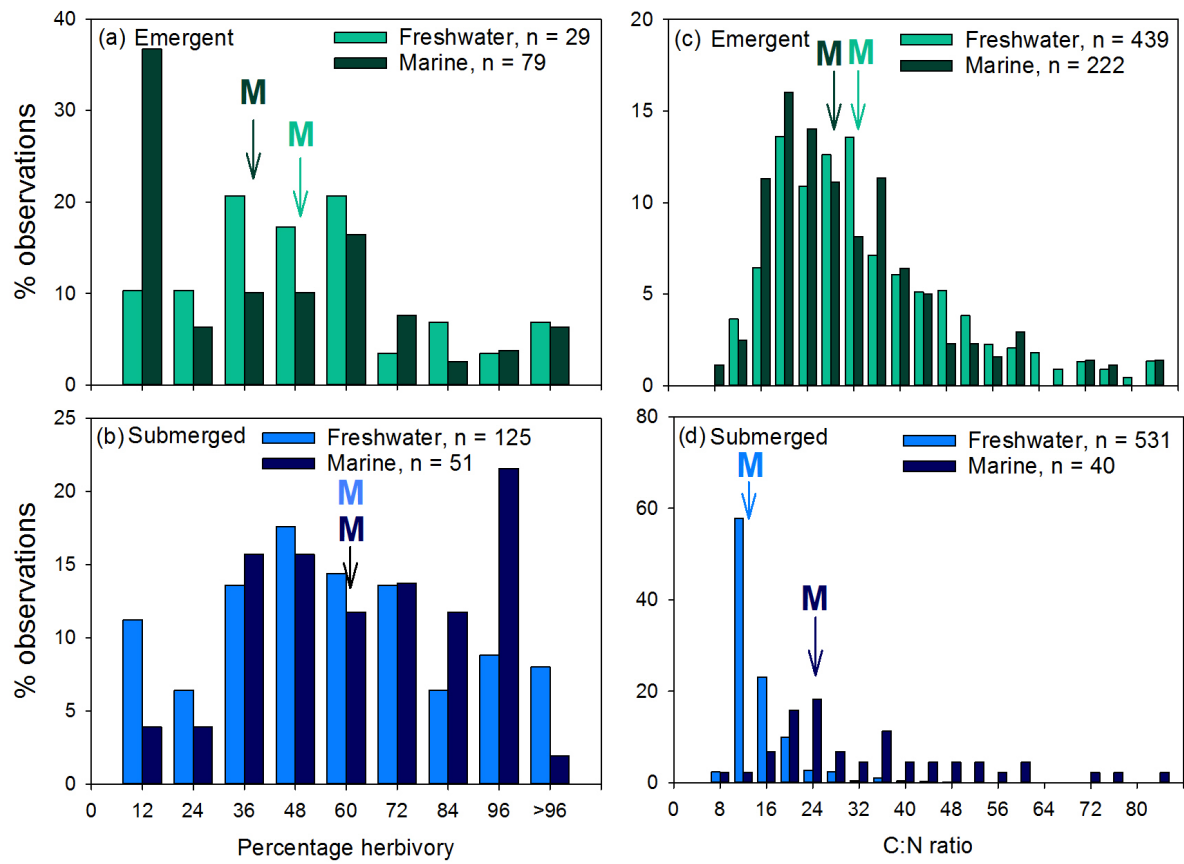
Natural gradients	Measure macrophyte performance (e.g. growth rate, biomass, etc) across naturally-occurring gradients in herbivore assemblage properties (e.g. density)	Wood et al. (2012b)
Drones	Identificaton of muskrat damage in constructed wetlands by digitizing low-altitude aerial photographs	Brandt et al. (2015)
Molecular markers	Assessments of genetic variation in plants across environmental or geographical (latitudinal) gradients using different molecular markers	Mader et al. (1998); Hangelbroek et al. (2002) ; King et al. (2002); Hidding et al. (2014)
Stable isotope analyses	Measurement of carbon, nitrogen and hydrogen stable isotopes in resources and consumers and application of mixing models	France et al. (1996); Solomon et al. (2011); Dorenbosch and Bakker (2012); Mendonca et al. (2013); Vander Zanden et al. (2013); Scharnweber et al. (2014); De Kluijver et al. (2015)
Gut analyses and stable isotope analyses	Gut analyses in fish combined with stable isotope analyses of basal food resources and fish	Mao et al. (2014)

Telemetry	Whole-lake fish telemetry	Hanson et al. (2007)
Bird counting and determination of lignin content in faeces		Dos Santos et al. (2012)
Laboratory feeding rate determination in fish	Determination of fish feeding rates in aquaria	Körner & Dugdale (2003)
Mechanistic models	Simulations of foraging herbivores or effects on plant growth can predict the location, timing, and magnitude of herbivore effects on macrophytes	Hootsmans (1999); Van Nes et al. (2003); Nolet et al. (2006); Wood et al. (2014a); Nolet et al. (2016); Hidding et al. (2016)
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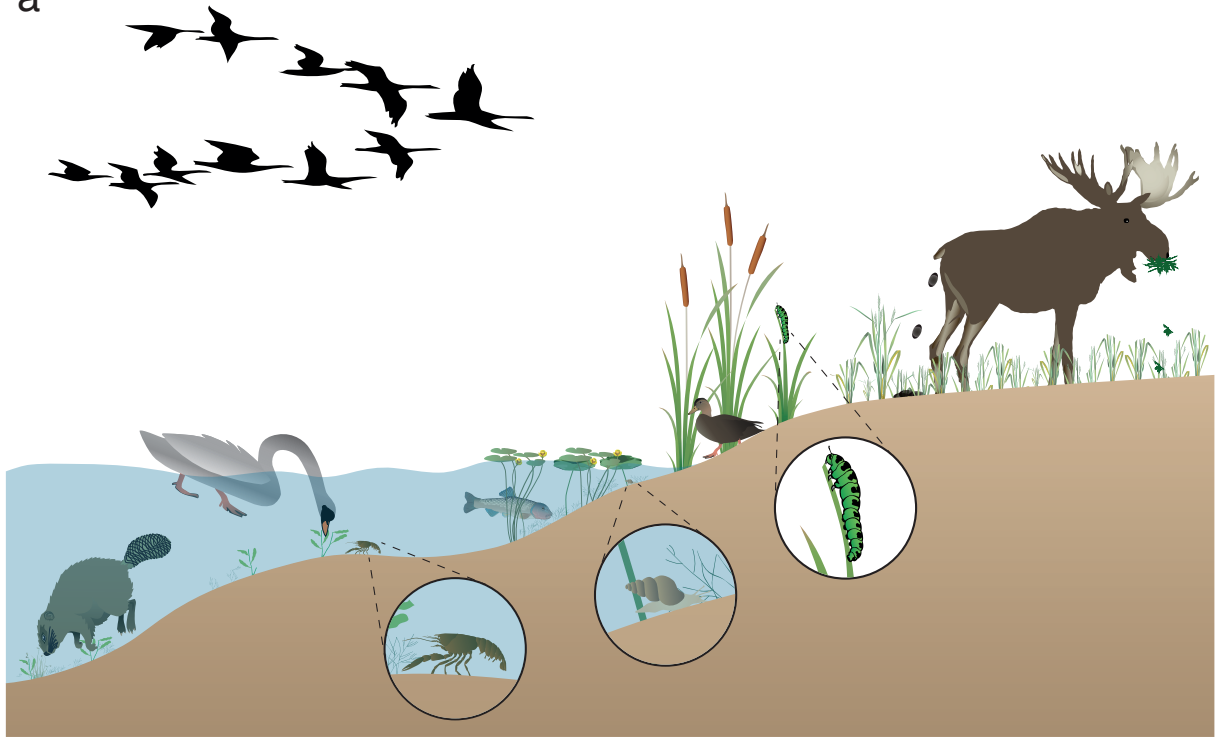
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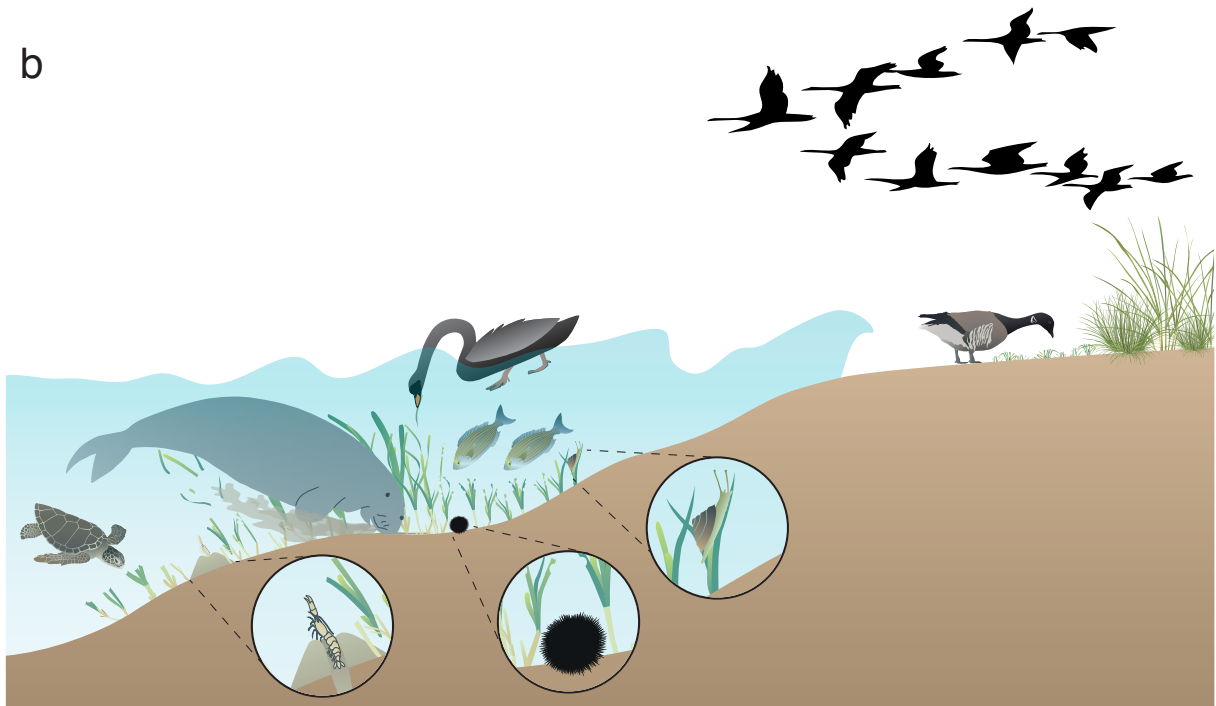
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